

Mathematical models in evolutionary research

Maria E. Orive

Ecology and Evolutionary Biology

University of Kansas

@MEOrive

NABT Professional Development Conference

November 10, 2018



Scott Williamson

mathematical models in ecology and evolution

Journal (2001)	Number of articles	Models used generally ¹	Models used specifically ²	Equations! ³
<i>American Naturalist</i>	105	96%	59%	58%
<i>Ecology</i>	274	100%	35%	38%
<i>Evolution</i>	231	100%	35%	33%

¹General use: includes statistical or phylogenetic analysis with a mathematical basis, e.g. ANOVA, regression, etc.

²Specific use: mathematical model used to obtain results

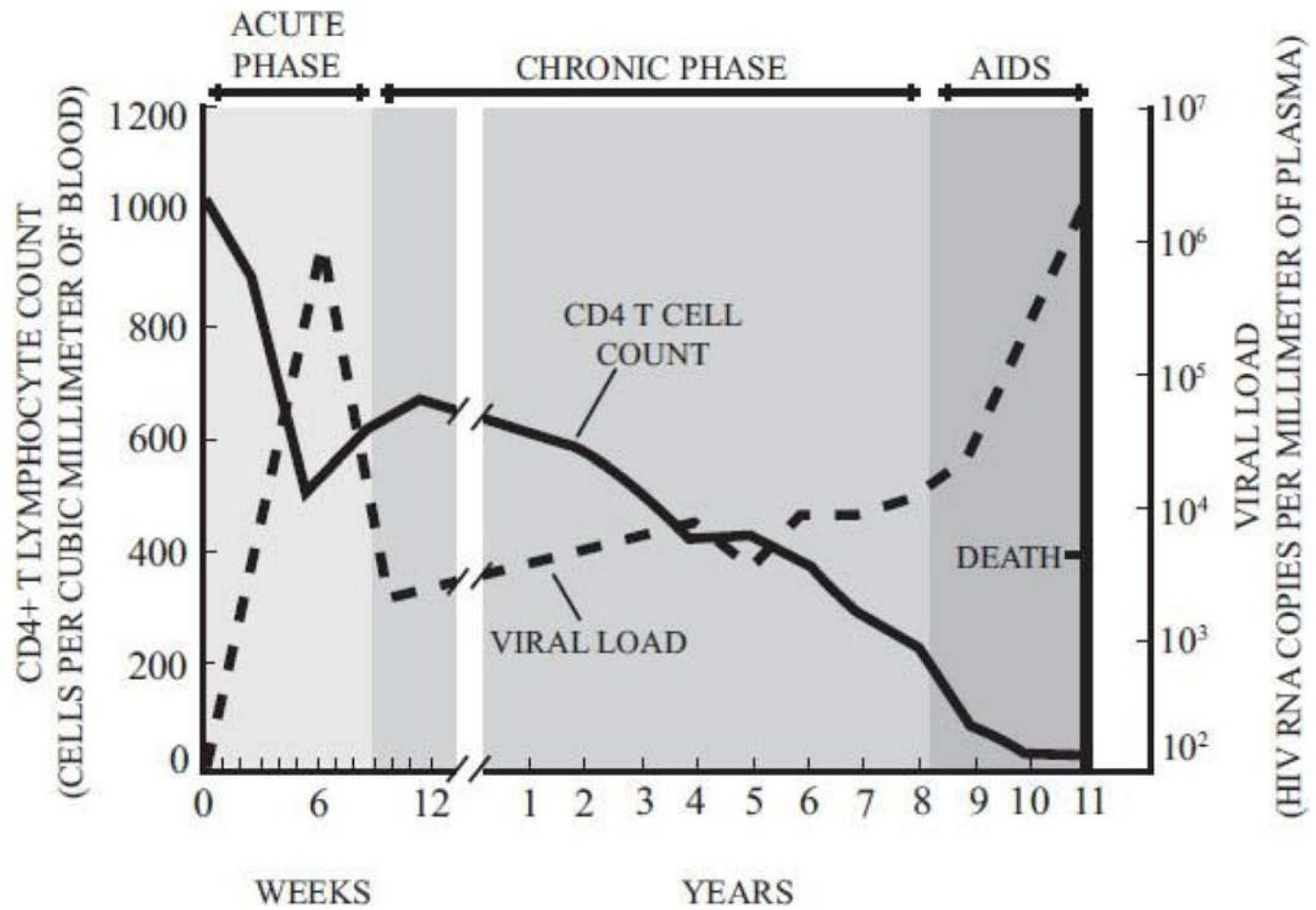
³Equations present: excluding standard statistical equations

adapted from Otto and Day (2007)

two ways we can use models to make sense of biology

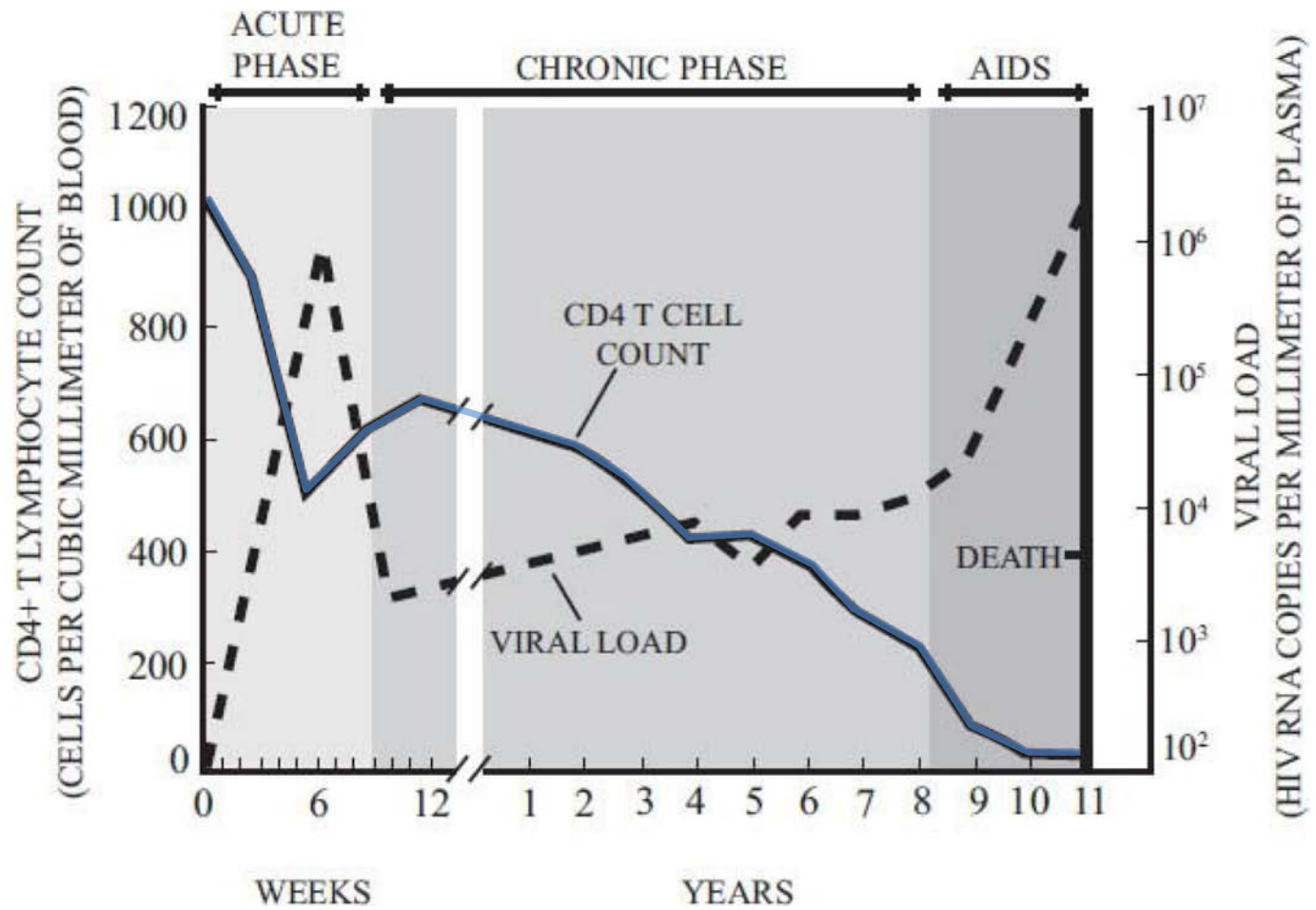
- Explain what we *do see*
 - Specific test of hypotheses
 - Example: dynamics of HIV after infection
- Predict what we *might see*
 - Generate hypotheses
 - Example: evolutionary lag and rescue with complex life histories

Time course of HIV infection within an individual



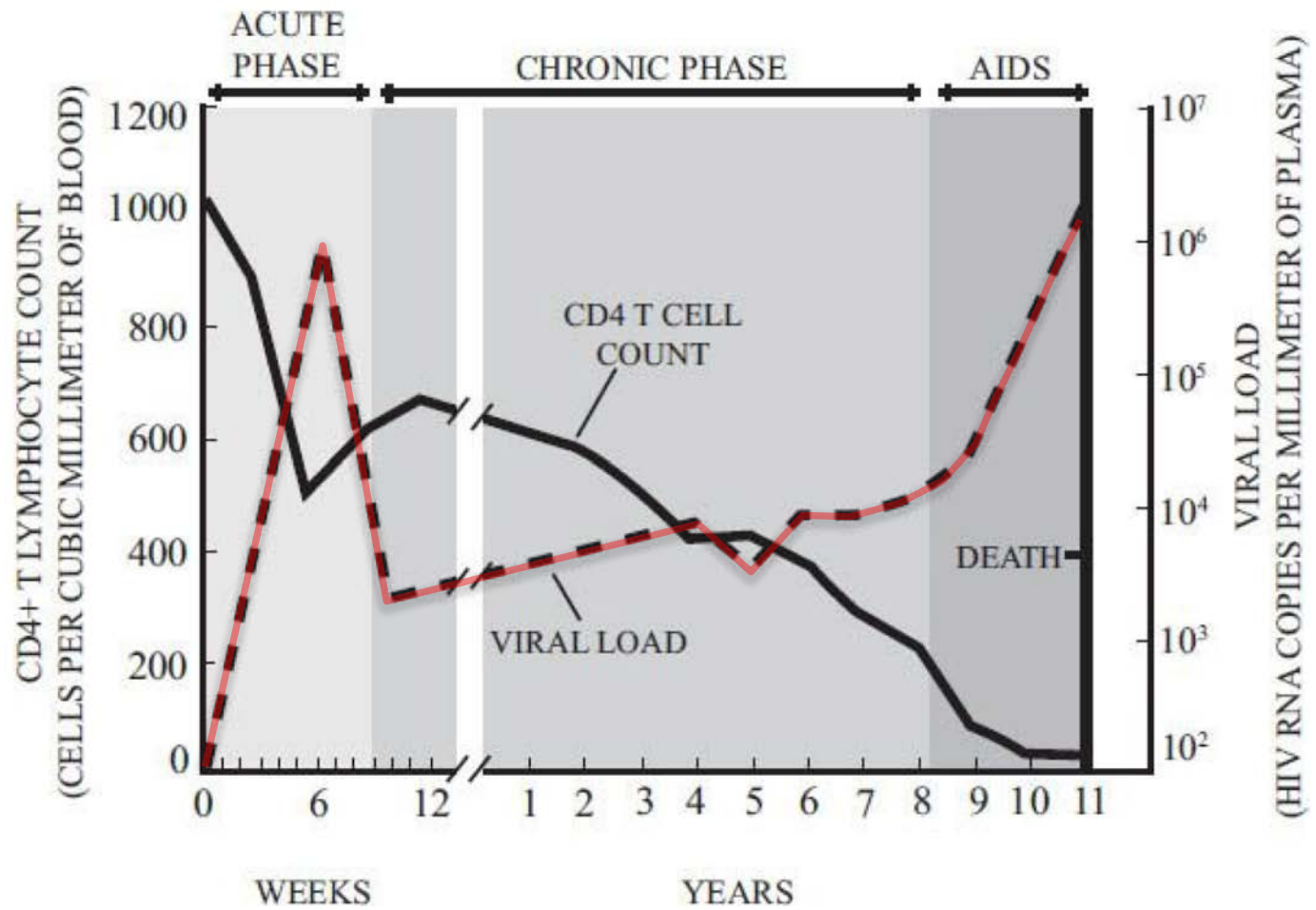
from Otto & Day (2007)
data from Fauci et al. (1996)

Time course of HIV infection within an individual



from Otto & Day (2007)
data from Fauci et al. (1996)

Time course of HIV infection within an individual



from Otto & Day (2007)
data from Fauci et al. (1996)

dynamical models: describing systems that change over time

differential equations – describe the *rate* at which
a variable changes over time;
continuous in time

$$\frac{d n(t)}{dt} = \text{"some function of } n(t)\text{"}$$

recursion equations – describe the *value* of a variable
in the next time step;
discrete in time

$$n(t + 1) = \text{"some function of } n(t)\text{"}$$

$$n' = \text{"some function of } n\text{"}$$

dynamical models: describing systems that change over time

differential equations – describe the *rate* at which
a variable changes over time;
continuous in time

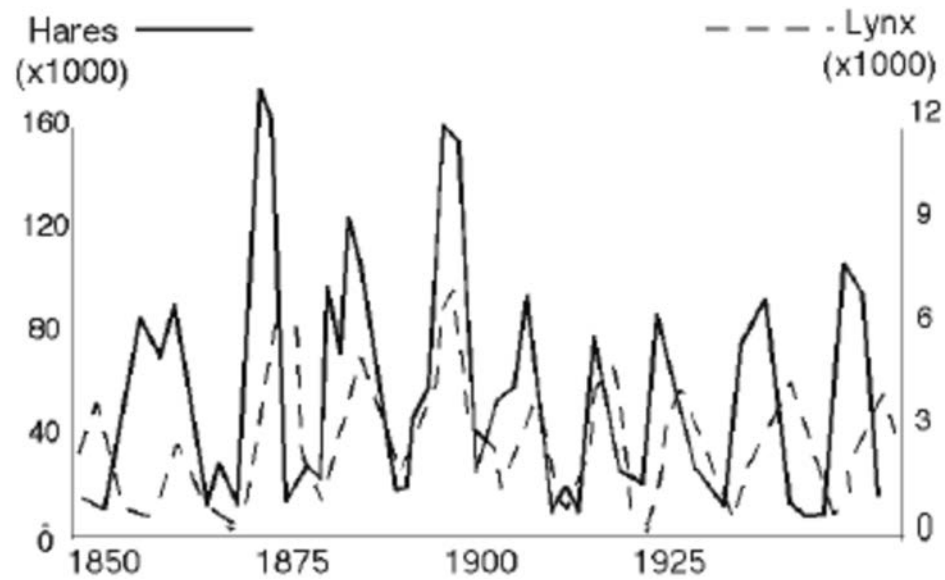
$$\frac{d n(t)}{dt} = \text{"some function of } n(t)\text{"}$$

recursion equations – describe the *value* of a variable
in the next time step;
discrete in time

$$n(t + 1) = \text{"some function of } n(t)\text{"}$$

$$n' = \text{"some function of } n\text{"}$$

example: predator-prey model



example: predator-prey model



prey $\frac{d x(t)}{d t} = a x(t) - b x(t) y(t)$

predator $\frac{d y(t)}{d t} = -c y(t) + p x(t) y(t)$

example: predator-prey model



prey $\frac{d x(t)}{dt} = a x(t) - b x(t) y(t)$

a = growth rate of prey (hares)

example: predator-prey model



prey
$$\frac{d x(t)}{d t} = a x(t) - b x(t) y(t)$$

b = capture rate (death rate of prey)

example: predator-prey model



p = growth rate of predator (lynx)

predator $\frac{d y(t)}{dt} = p x(t) y(t) - c y(t)$

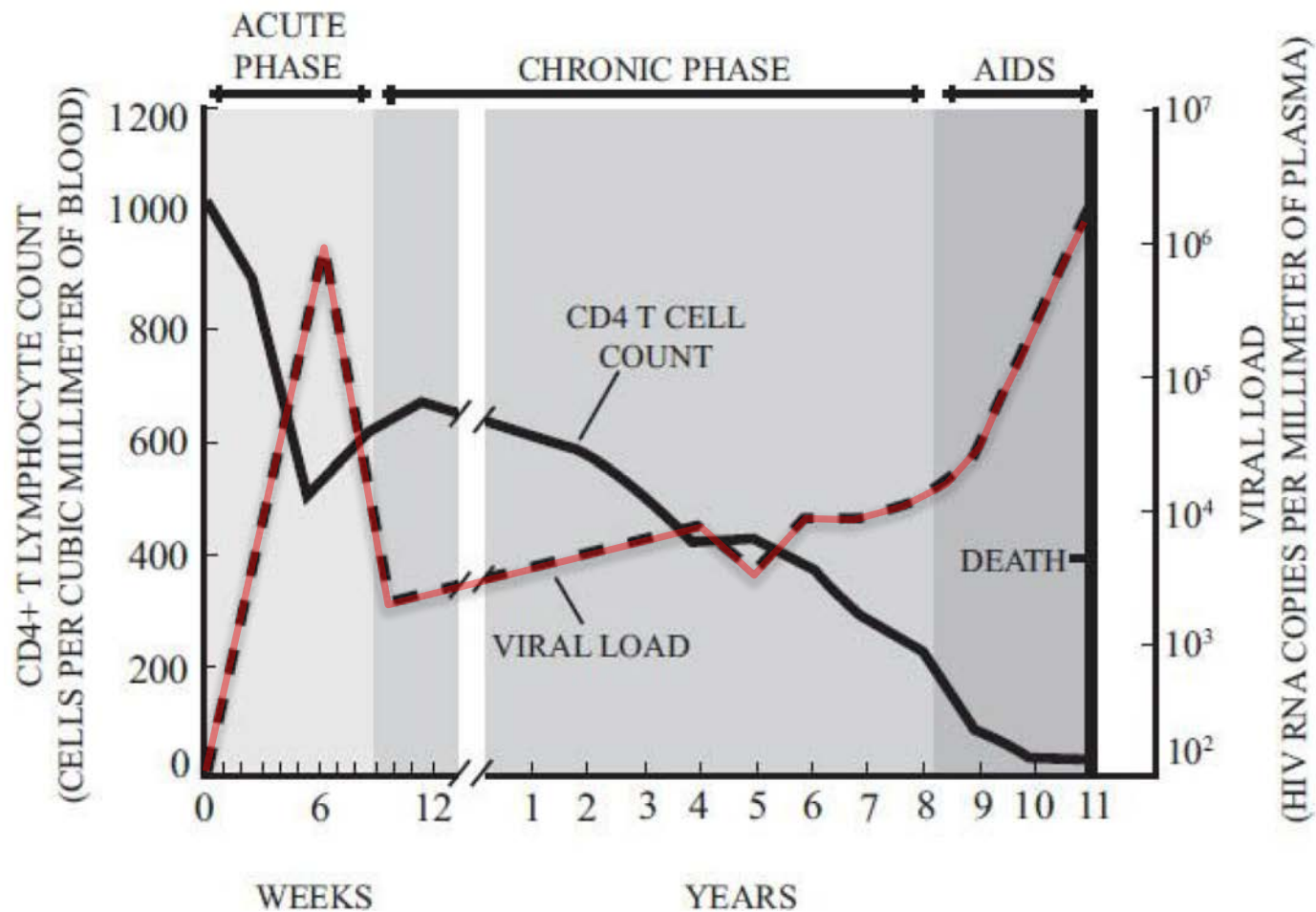
example: predator-prey model



c = death rate of predator (lynx)

predator $\frac{d y(t)}{dt} = p x(t) y(t) - c y(t)$

Time course of HIV infection within an individual



from Otto & Day (2007)
data from Fauci et al. (1996)

model of within-individual HIV infection (Phillips 1996)

susceptible CD4+ cells $\frac{dR(t)}{dt} = \Gamma \tau - \mu R(t) - \beta V(t) R(t)$

latently infected CD4+ cells $\frac{dL(t)}{dt} = p \beta V(t) R(t) - \mu L(t) - \alpha L(t)$

actively infected CD4+ cells $\frac{dE(t)}{dt} = (1 - p) \beta V(t) R(t) + \alpha L(t) - \delta E(t)$

virus particles $\frac{dV(t)}{dt} = \pi E(t) - \sigma V(t) - \beta V(t) R(t)$

model of within-individual HIV infection (Phillips 1996)

susceptible CD4+ cells $\frac{dR(t)}{dt} = \Gamma \tau - \mu R(t) - \beta V(t) R(t)$

latently infected CD4+ cells $\frac{dL(t)}{dt} = p \beta V(t) R(t) - \mu L(t) - \alpha L(t)$

actively infected CD4+ cells $\frac{dE(t)}{dt} = (1 - p) \beta V(t) R(t) + \alpha L(t) - \delta E(t)$

predator

virus particles $\frac{dV(t)}{dt} = \pi E(t) - \sigma V(t) - \beta V(t) R(t)$

model of within-individual HIV infection (Phillips 1996)

prey

susceptible CD4+ cells

$$\frac{dR(t)}{dt} = \Gamma \tau - \mu R(t) - \beta V(t) R(t)$$

latently infected CD4+ cells

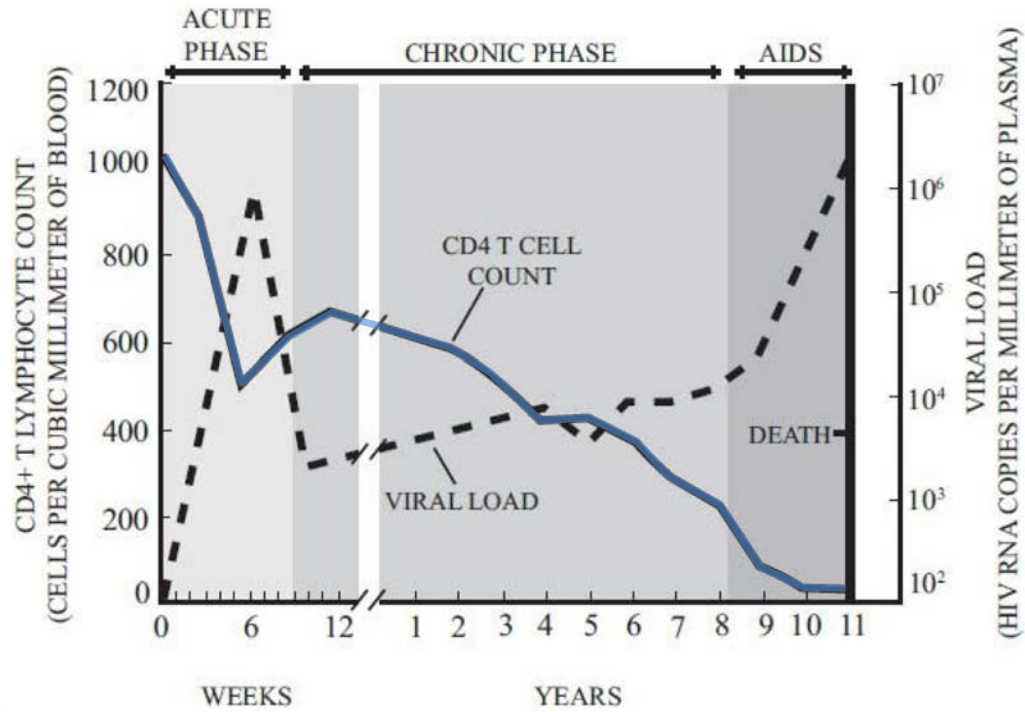
$$\frac{dL(t)}{dt} = p \beta V(t) R(t) - \mu L(t) - \alpha L(t)$$

actively infected CD4+ cells

$$\frac{dE(t)}{dt} = (1 - p) \beta V(t) R(t) + \alpha L(t) - \delta E(t)$$

virus particles

$$\frac{dV(t)}{dt} = \pi E(t) - \sigma V(t) - \beta V(t) R(t)$$

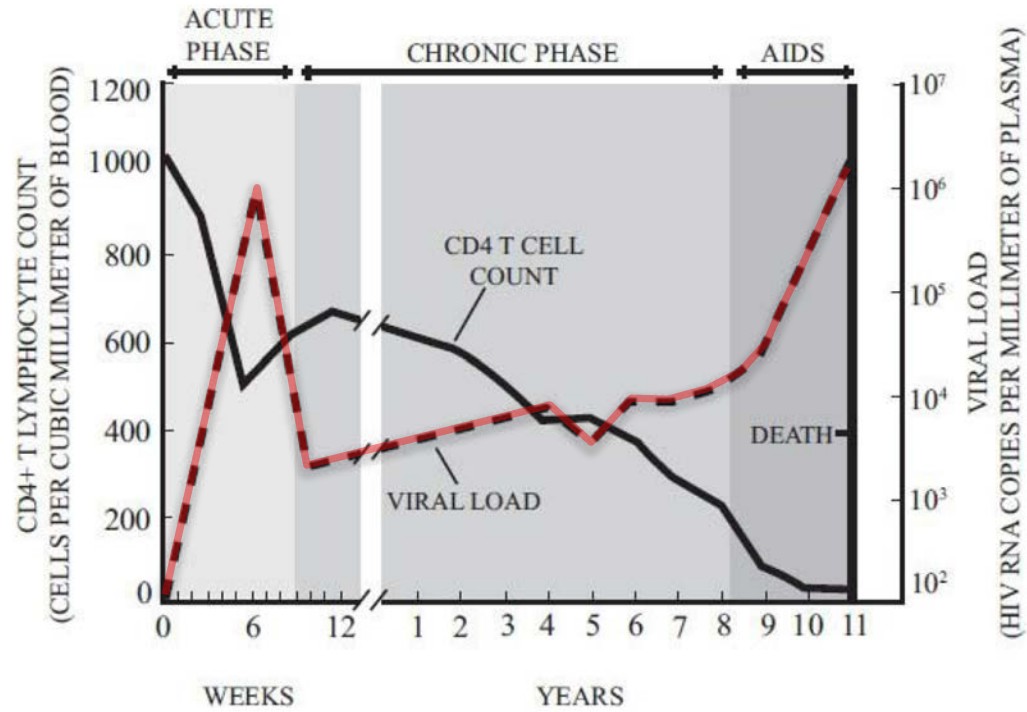


number of CD4+ cells
(susceptible + latent + active)

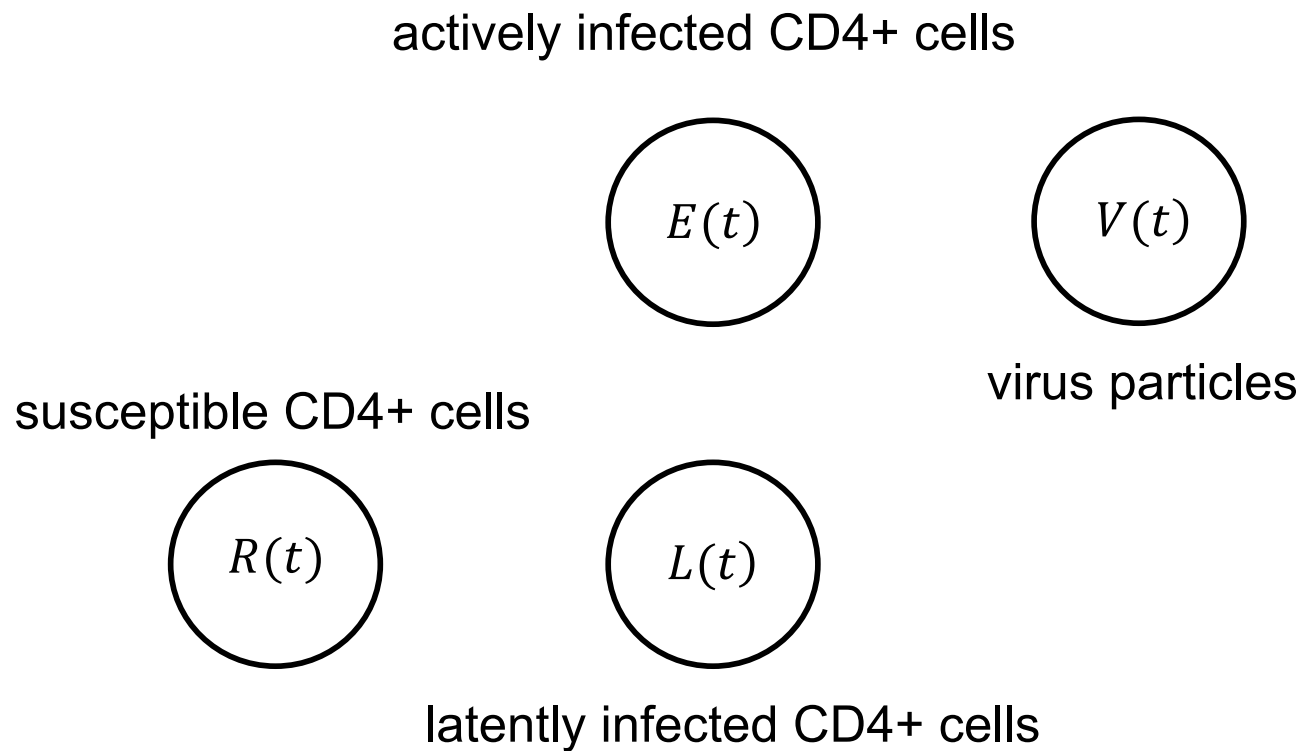
$$\frac{dR(t)}{dt} = \Gamma \tau - \mu R(t) - \beta V(t) R(t)$$

$$\frac{dL(t)}{dt} = p \beta V(t) R(t) - \mu L(t) - \alpha L(t)$$

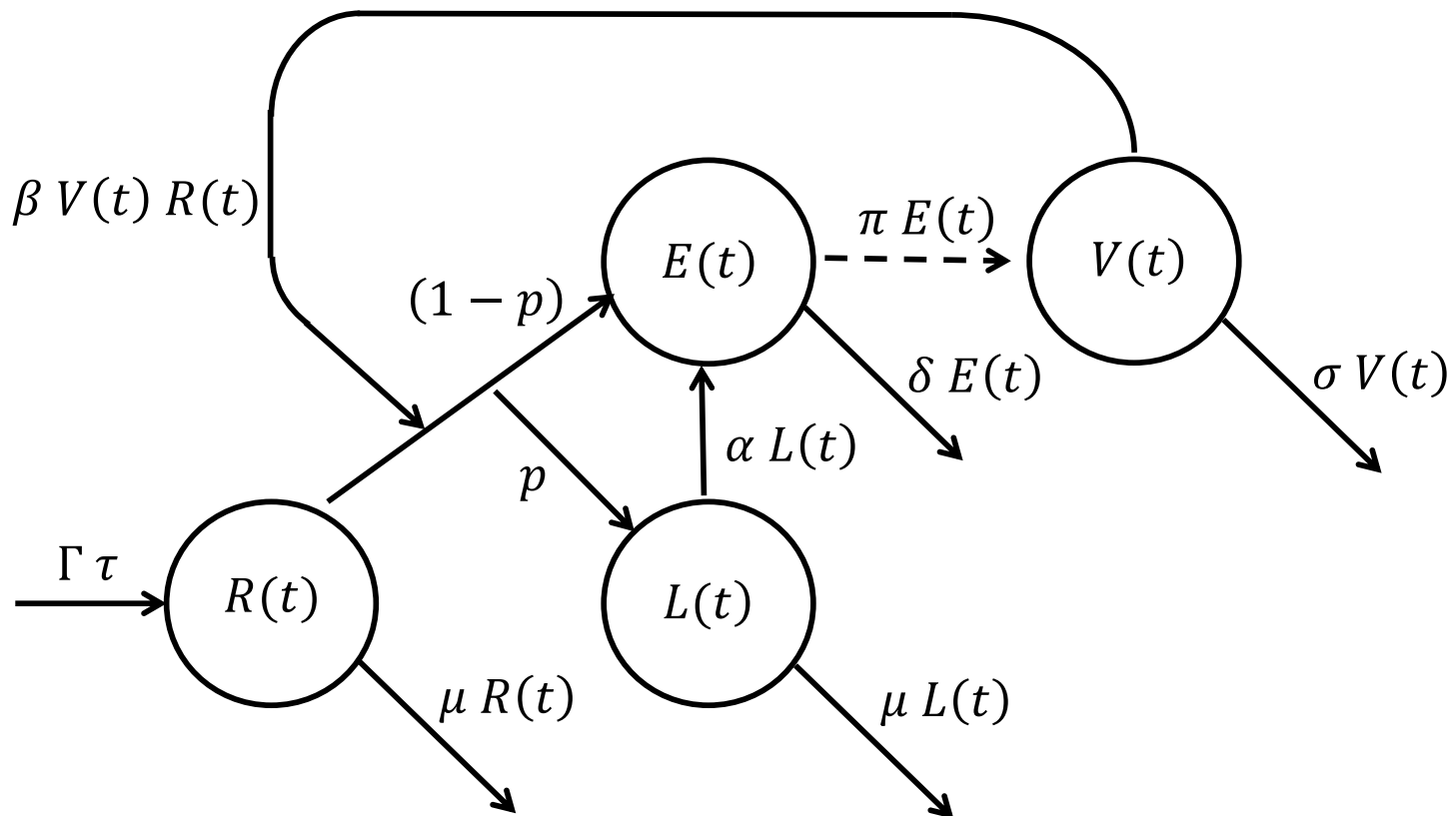
$$\frac{dE(t)}{dt} = (1 - p) \beta V(t) R(t) + \alpha L(t) - \delta E(t)$$



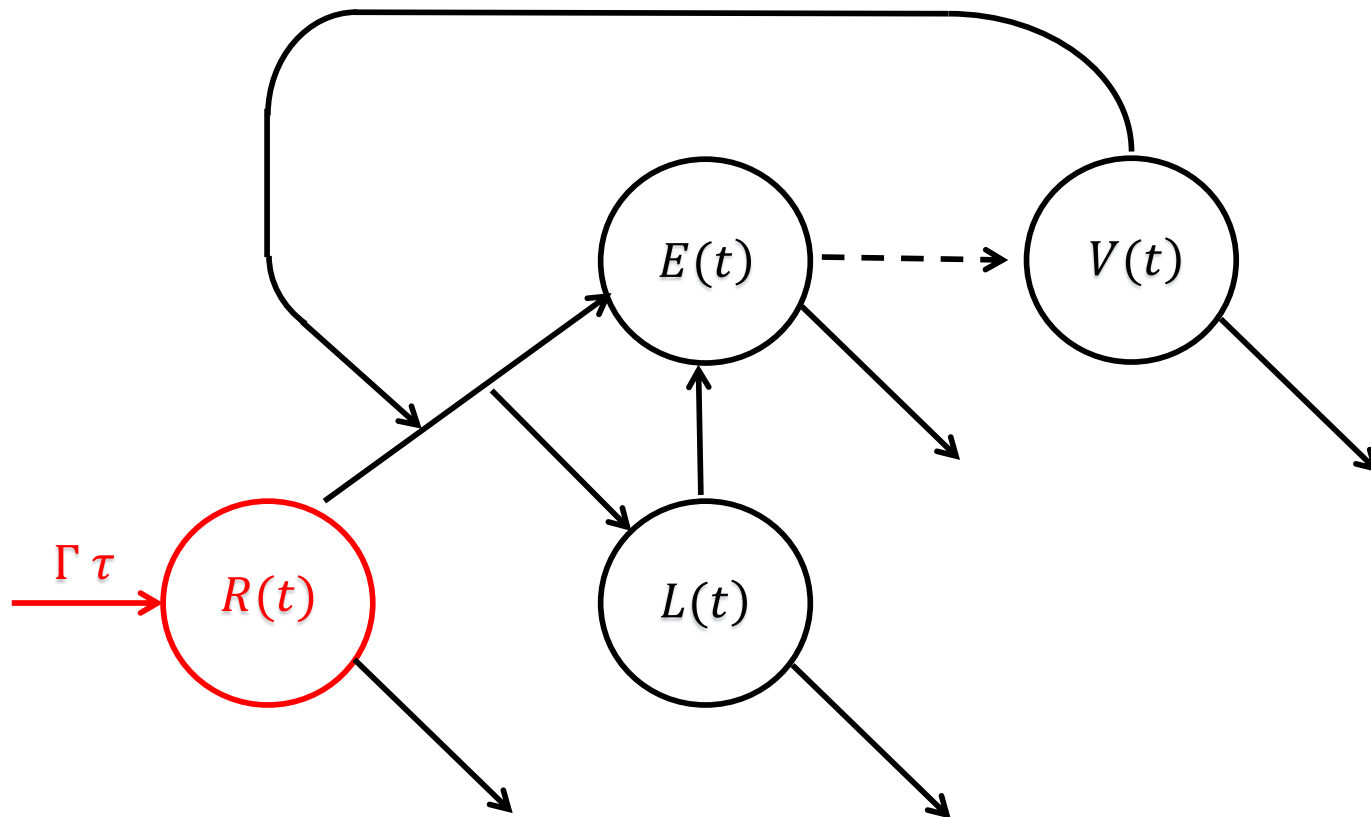
number of virus particles $\left\{ \begin{array}{l} \frac{dV(t)}{dt} = \pi E(t) - \sigma V(t) - \beta V(t) R(t) \end{array} \right.$



Flow diagram for Phillips (1996) model
adapted from Otto & Day (2007)



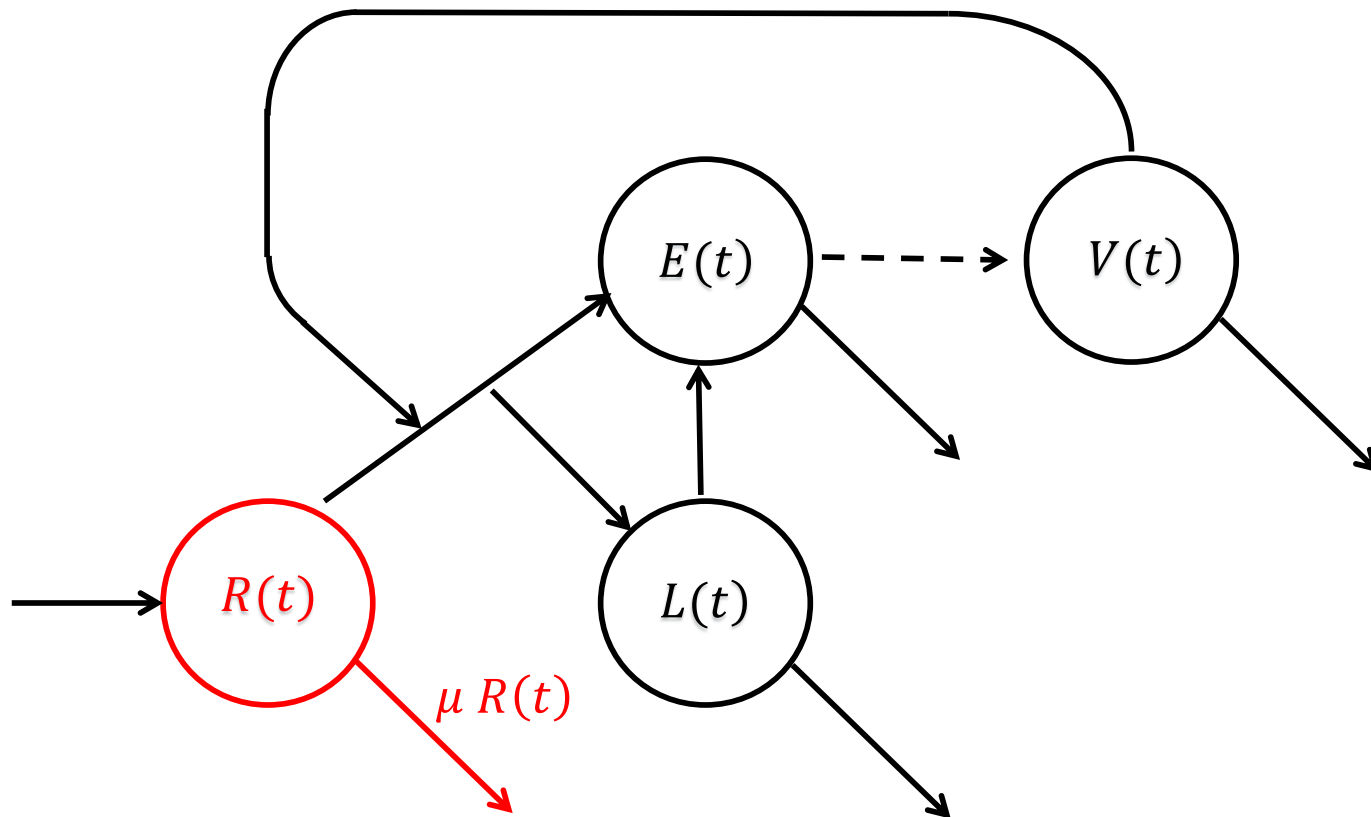
Flow diagram for Phillips (1996) model adapted from Otto & Day (2007)



susceptible CD4+ cells

$$\frac{dR(t)}{dt} = \Gamma \tau - \mu R(t) - \beta V(t) R(t)$$

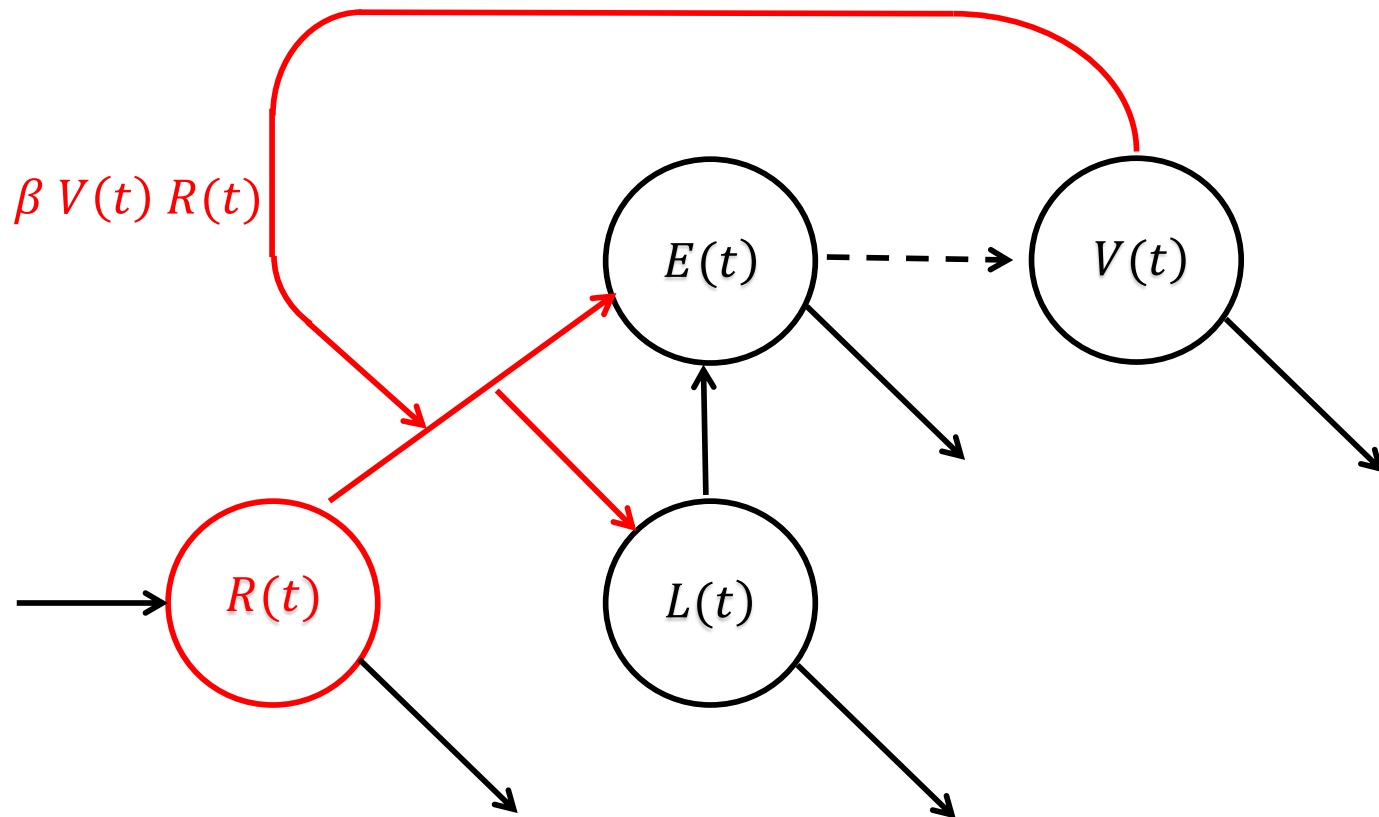
$\Gamma \tau$ = input of susceptible cells from immune system



susceptible CD4+ cells

$$\frac{dR(t)}{dt} = \Gamma \tau - \mu R(t) - \beta V(t) R(t)$$

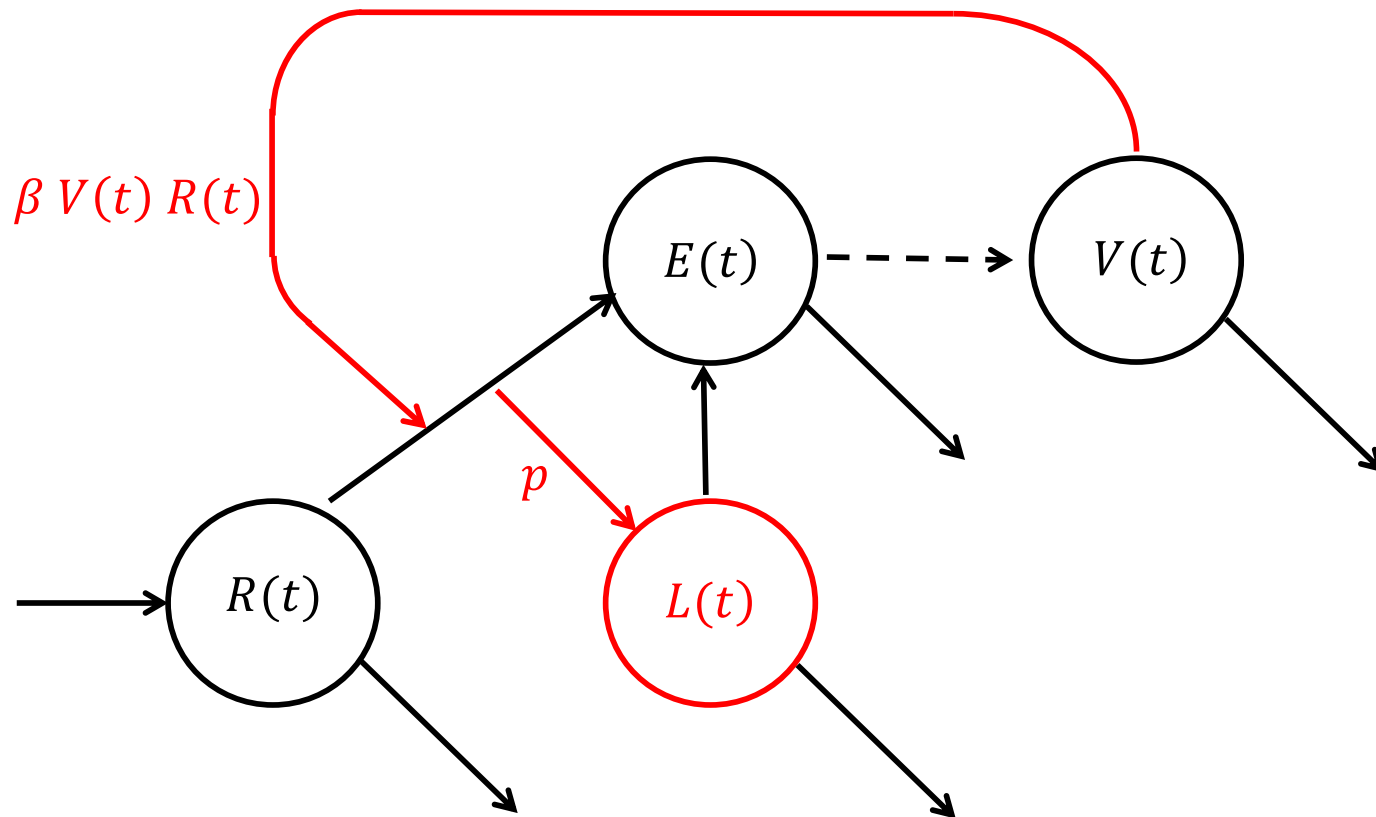
μ = death rate of CD4+ cells



susceptible CD4+ cells

$$\frac{dR(t)}{dt} = \Gamma \tau - \mu R(t) - \beta V(t) R(t)$$

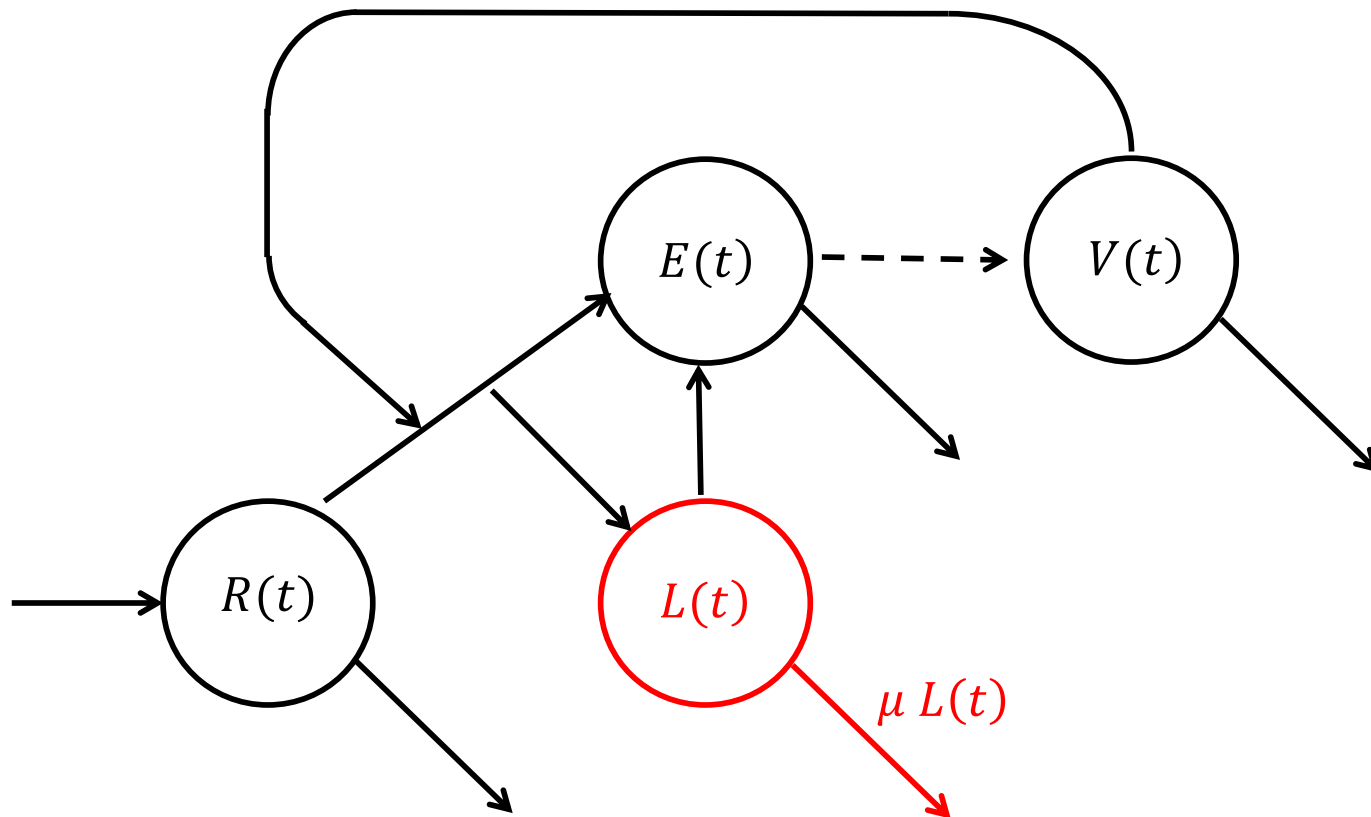
β = infection rate of susceptible cells



latently infected CD4+ cells

$$\frac{dL(t)}{dt} = p \beta V(t) R(t) - \mu L(t) - \alpha L(t)$$

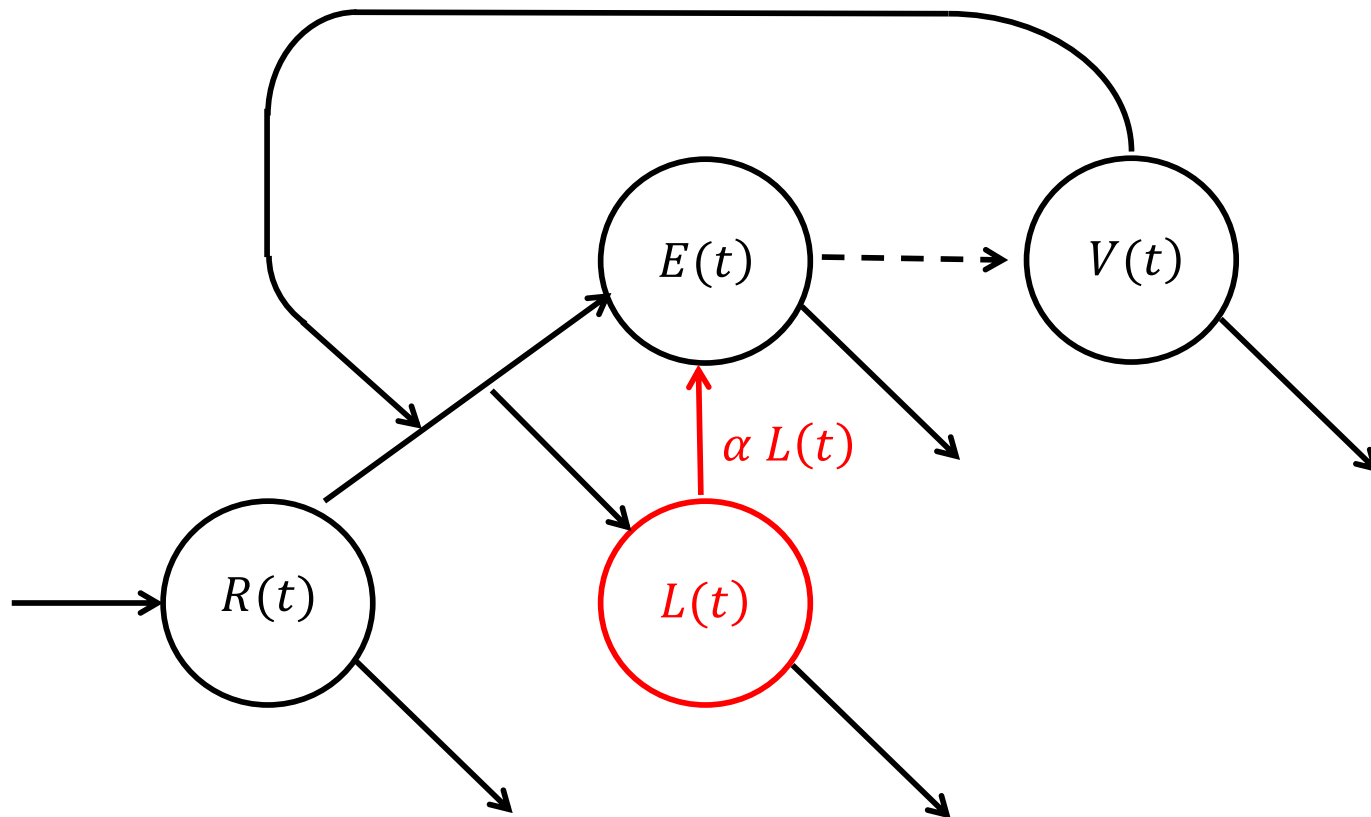
p = probability HIV in infected cell is latent



latently infected CD4+ cells

$$\frac{dL(t)}{dt} = p \beta V(t) R(t) - \mu L(t) - \alpha L(t)$$

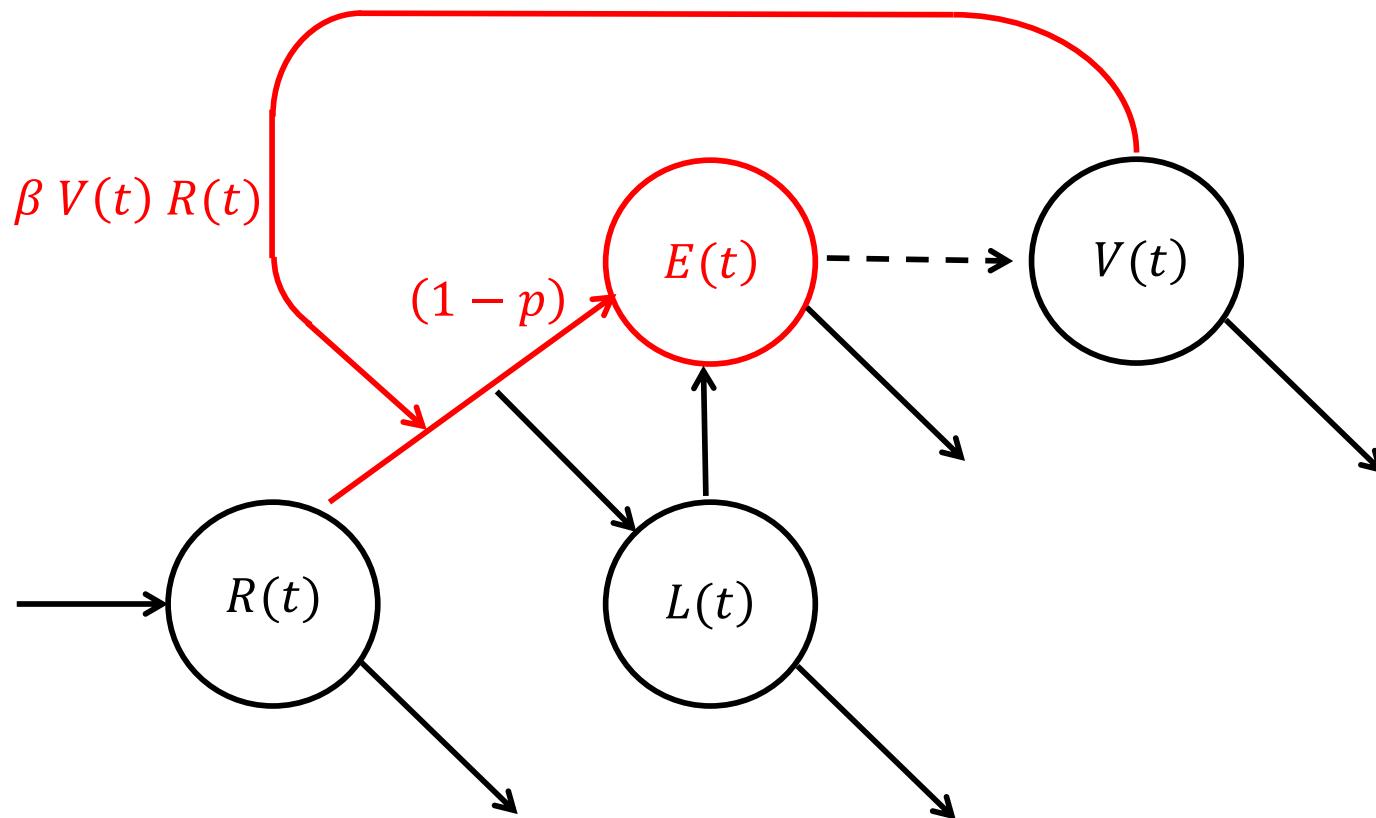
μ = death rate of CD4+ cells



latently infected CD4+ cells

$$\frac{dL(t)}{dt} = p \beta V(t) R(t) - \mu L(t) - \alpha L(t)$$

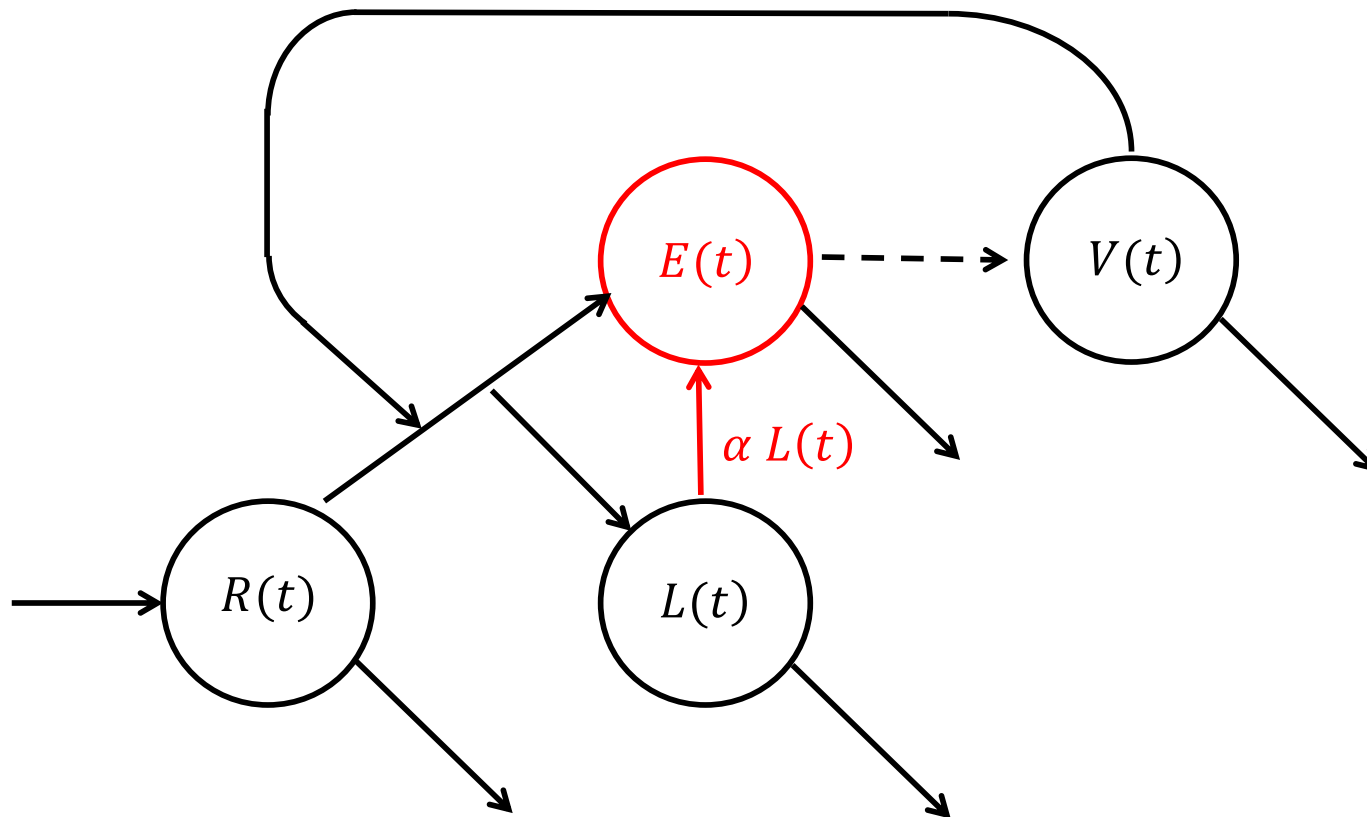
α = conversion rate from latent to active



actively infected CD4+ cells

$$\frac{dE(t)}{dt} = (1-p) \beta V(t) R(t) + \alpha L(t) - \delta E(t)$$

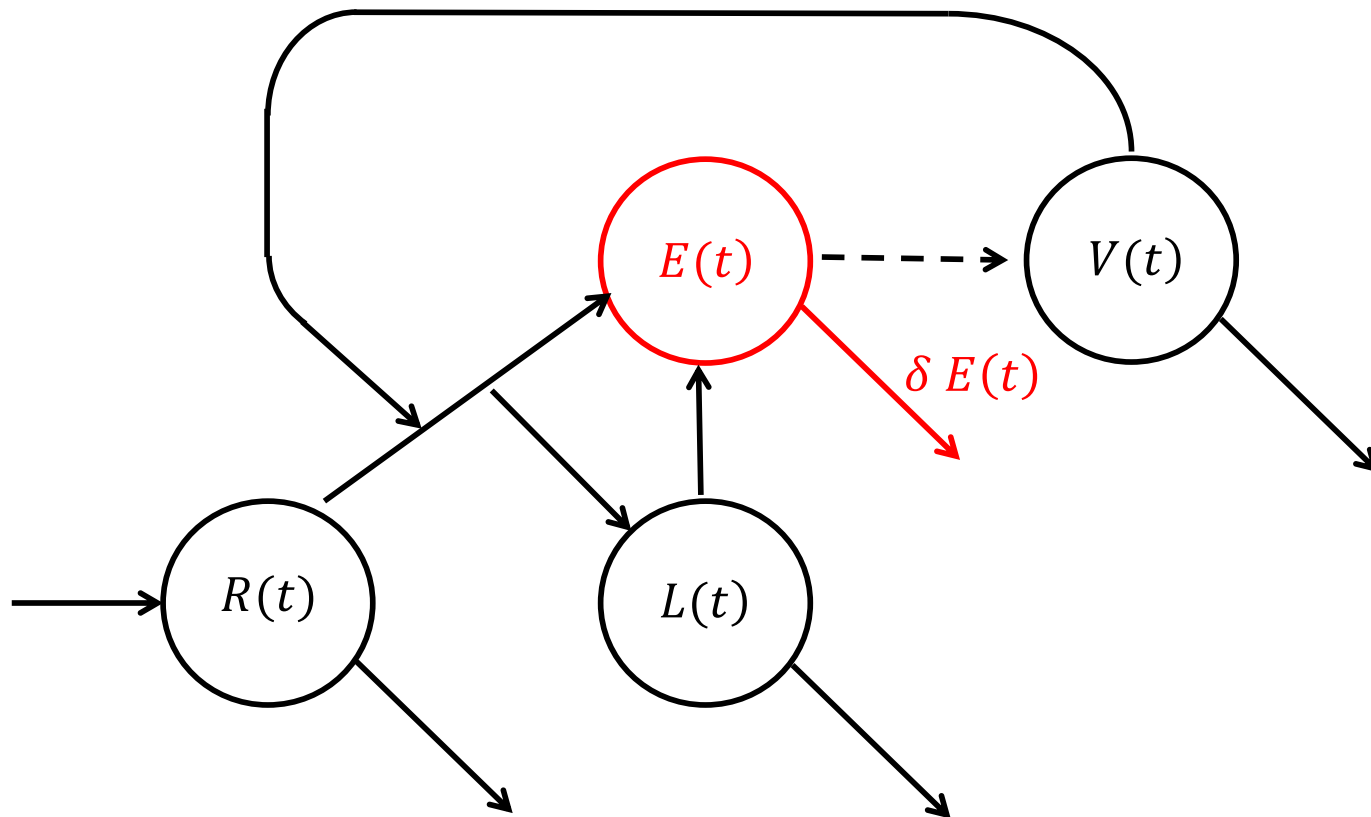
$1-p$ = probability HIV in infected cell is active



actively infected CD4+ cells

$$\frac{dE(t)}{dt} = (1 - p) \beta V(t) R(t) + \alpha L(t) - \delta E(t)$$

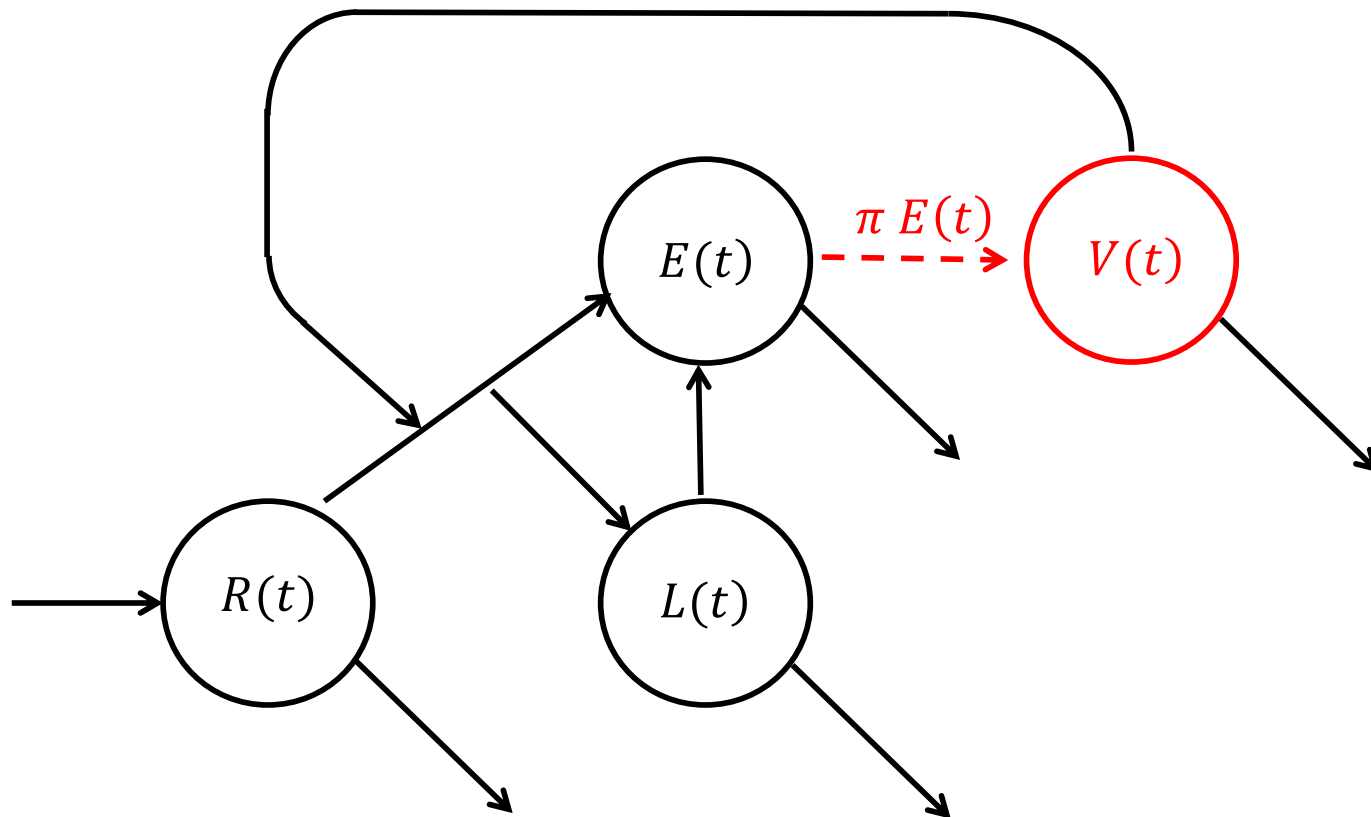
α = conversion rate from latent to active



actively infected CD4+ cells

$$\frac{dE(t)}{dt} = (1 - p) \beta V(t) R(t) + \alpha L(t) - \delta E(t)$$

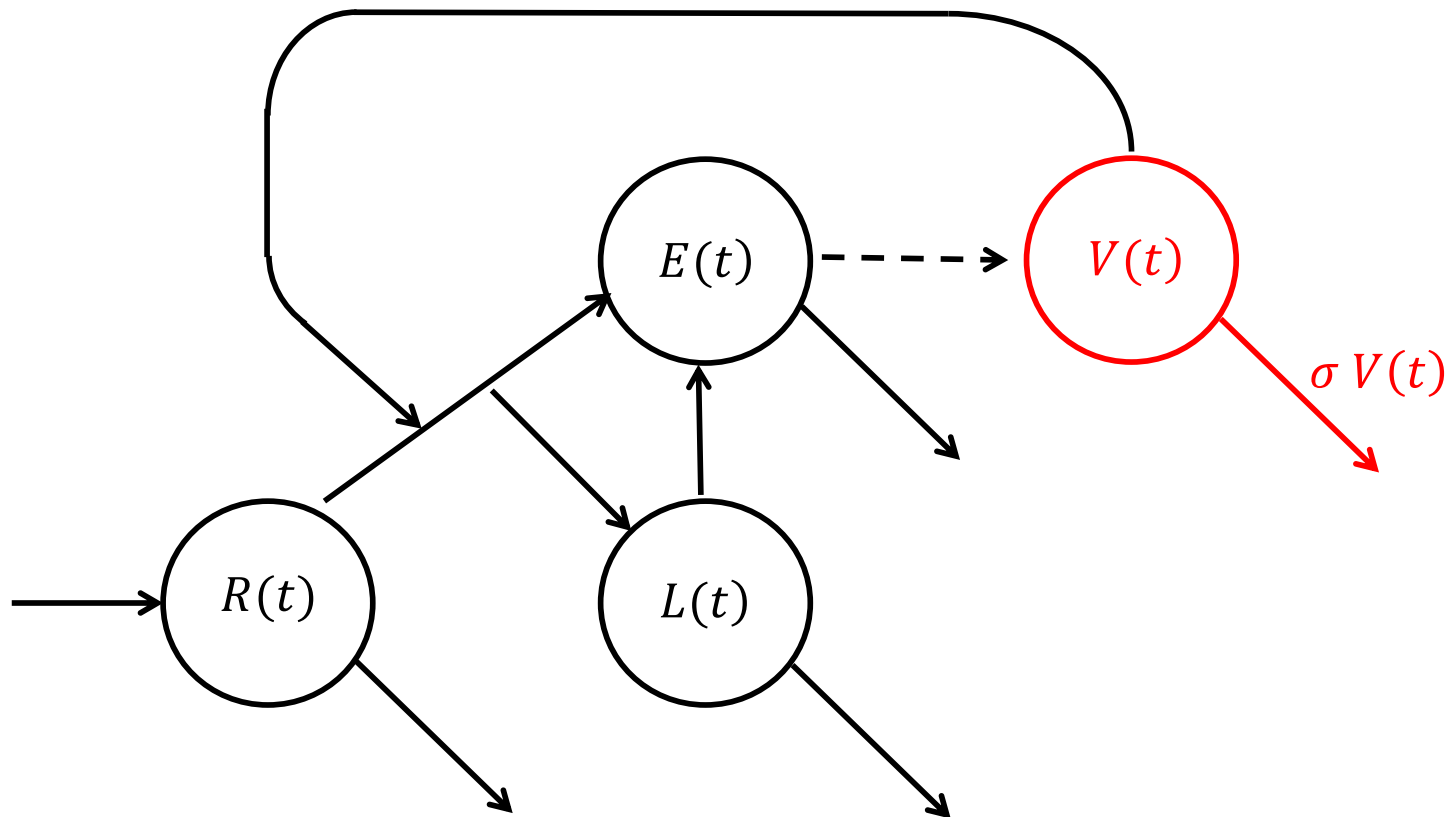
δ = death rate of actively infected CD4+ cells



virus particles

$$\frac{dV(t)}{dt} = \pi E(t) - \sigma V(t) - \beta V(t) R(t)$$

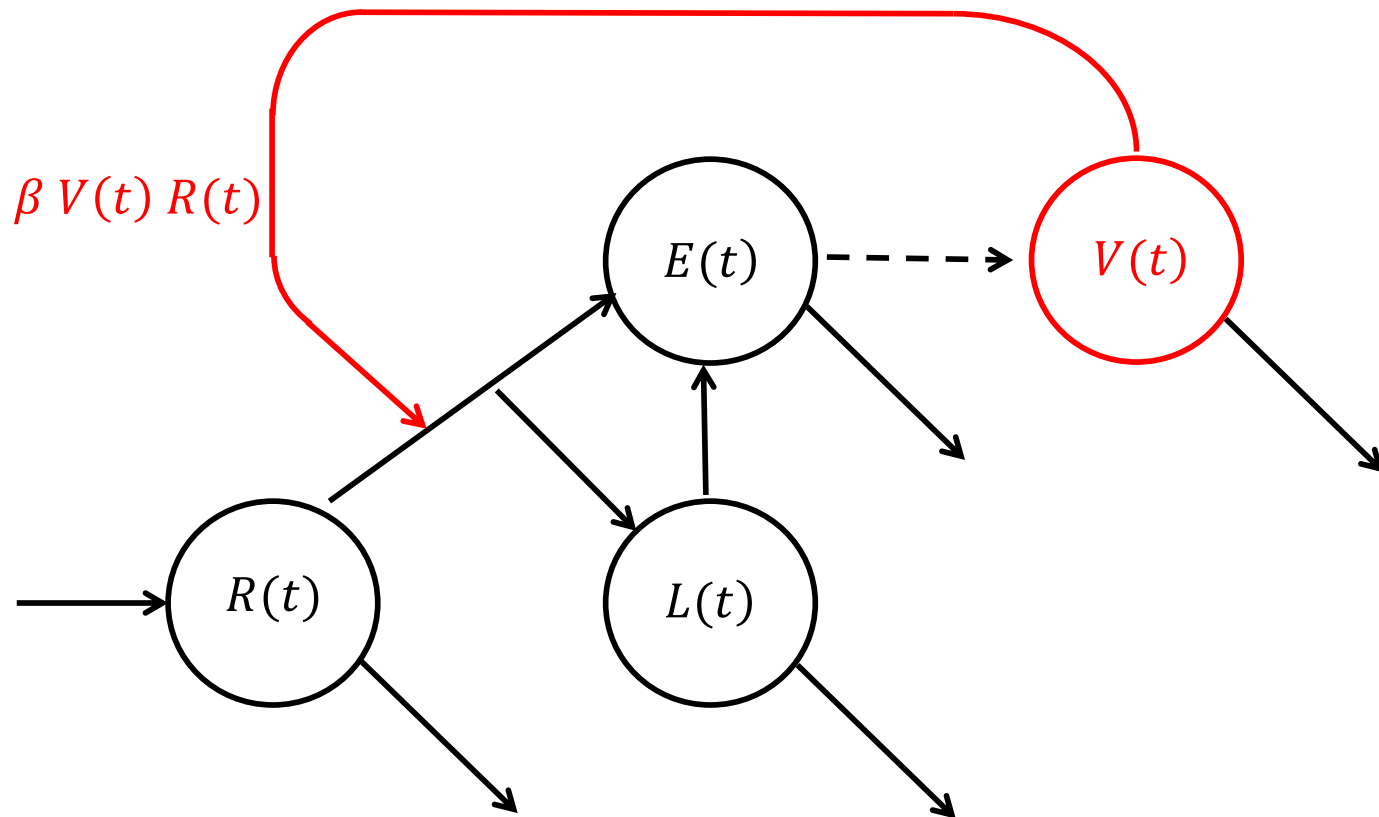
π = budding rate of virus particles from infected cells



virus particles

$$\frac{dV(t)}{dt} = \pi E(t) - \sigma V(t) - \beta V(t) R(t)$$

σ = clearance rate of virus particles



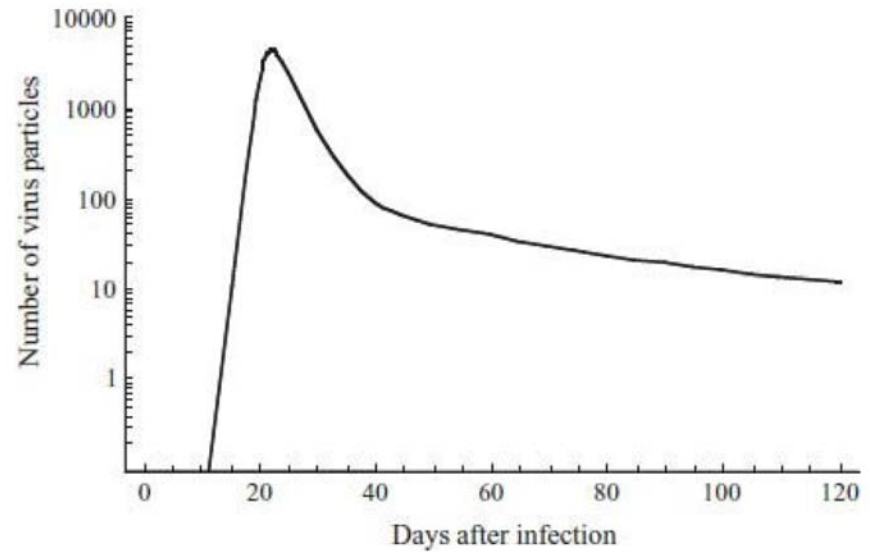
virus particles

$$\frac{dV(t)}{dt} = \pi E(t) - \sigma V(t) - \beta V(t) R(t)$$

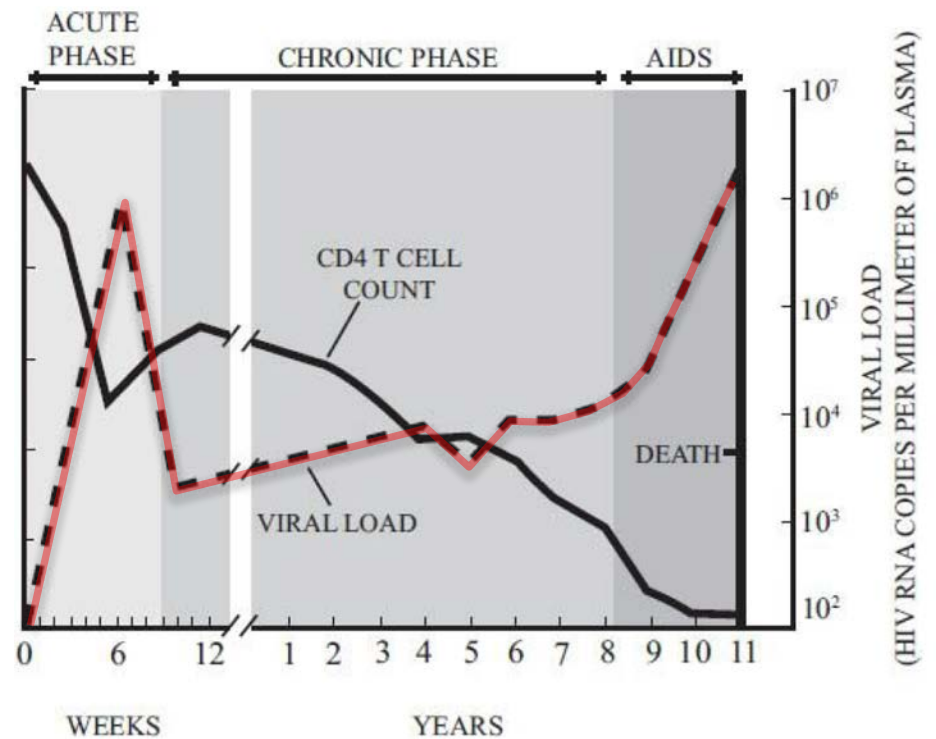
β = infection rate

Number of virus particles over time

Model: Phillips (1996)



Data: Fauci et al. (1996)



Linking Dynamical and Population Genetic Models of Persistent Viral Infection

John K. Kelly,^{1,*} Scott Williamson,¹ Maria E. Orive,¹ Marilyn S. Smith,² and Robert D. Holt³

Kelly, Williamson, et al. 2003 *Am Nat*

Adaptation in the *env* Gene of HIV-1 and Evolutionary Theories of Disease Progression

Scott Williamson

Department of Ecology and Evolutionary Biology, University of Kansas

Williamson 2003 *Mol Biol Evol*

A Statistical Characterization of Consistent Patterns of Human Immunodeficiency Virus Evolution Within Infected Patients

Scott Williamson,† Steven M. Perry,† Carlos D. Bustamante,* Maria E. Orive,† Miles N. Stearns,† and John K. Kelly†*

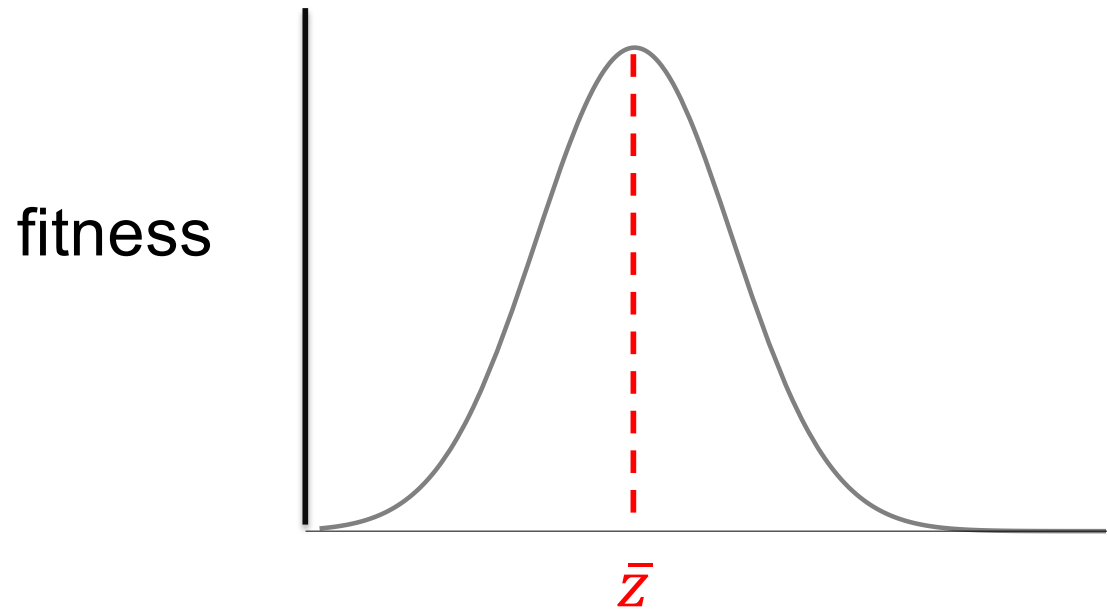
*Department of Biological Statistics and Computational Biology, Cornell University, Ithaca, New York; †Department of Ecology and Evolutionary Biology, University of Kansas, Lawrence

Williamson et al. 2005 *Mol Biol Evol*

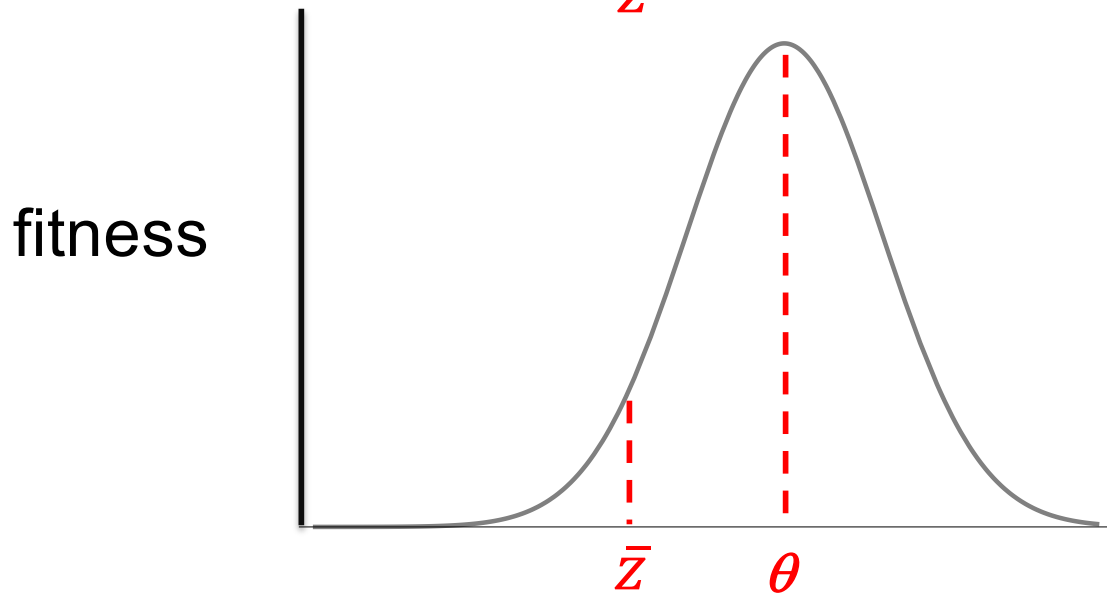
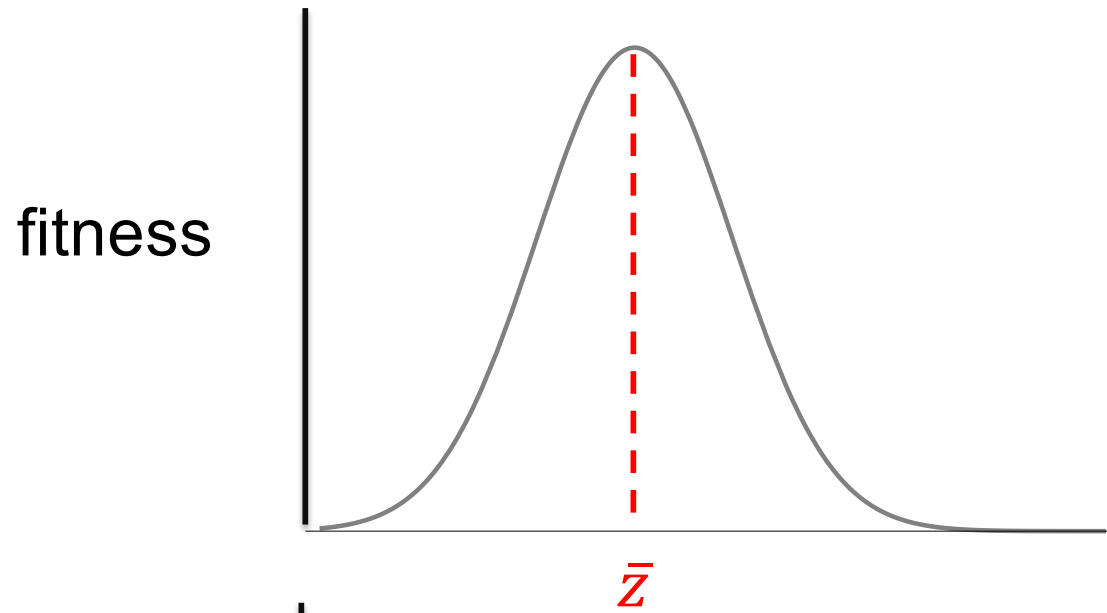
two ways we can use models to make sense of biology

- Explain what we *do see*
 - Specific test of hypotheses
 - Example: dynamics of HIV after infection
- Predict what we *might see*
 - Generate hypotheses
 - Example: evolutionary lag and rescue with complex life histories

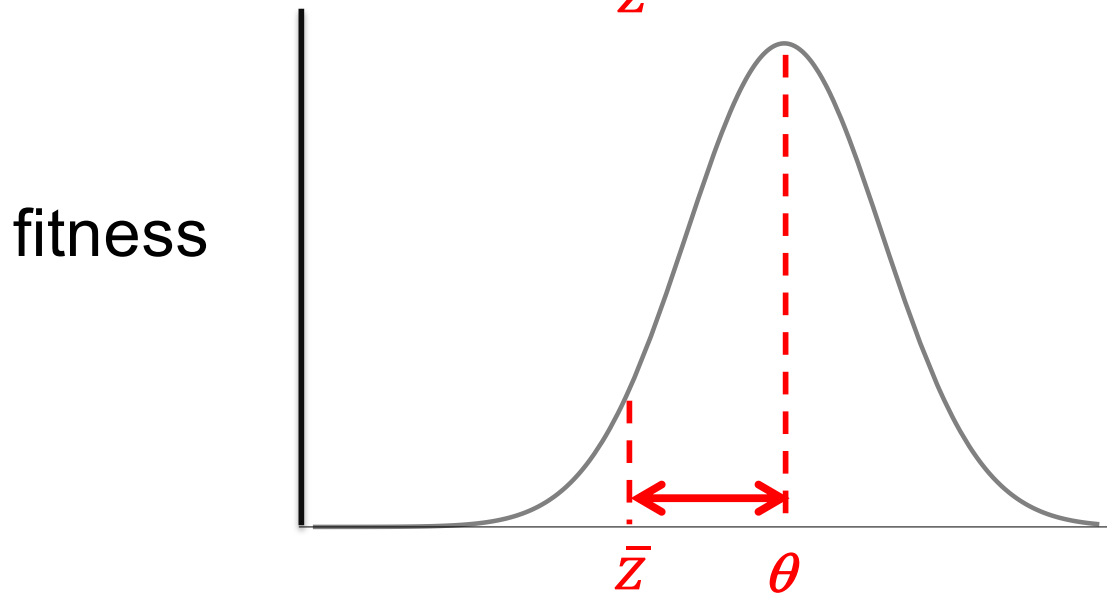
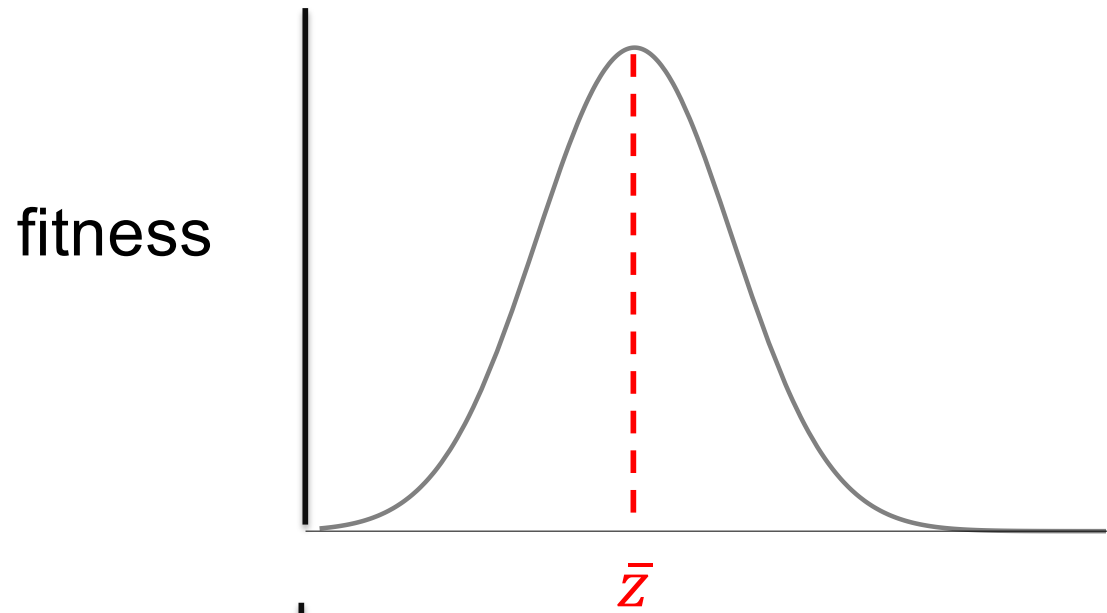
tracking environmental change



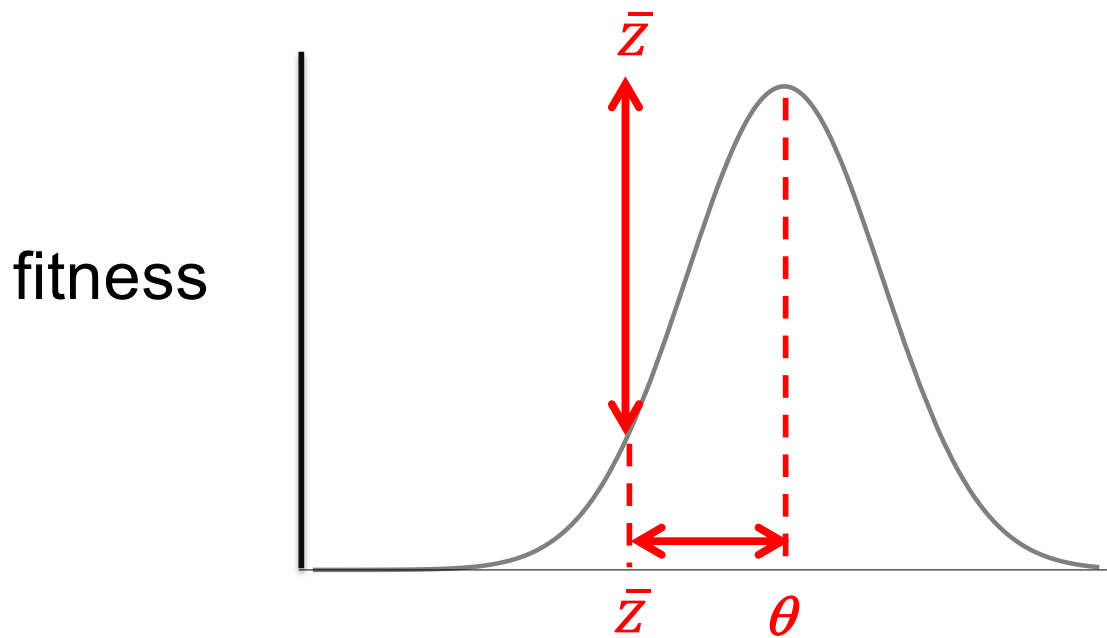
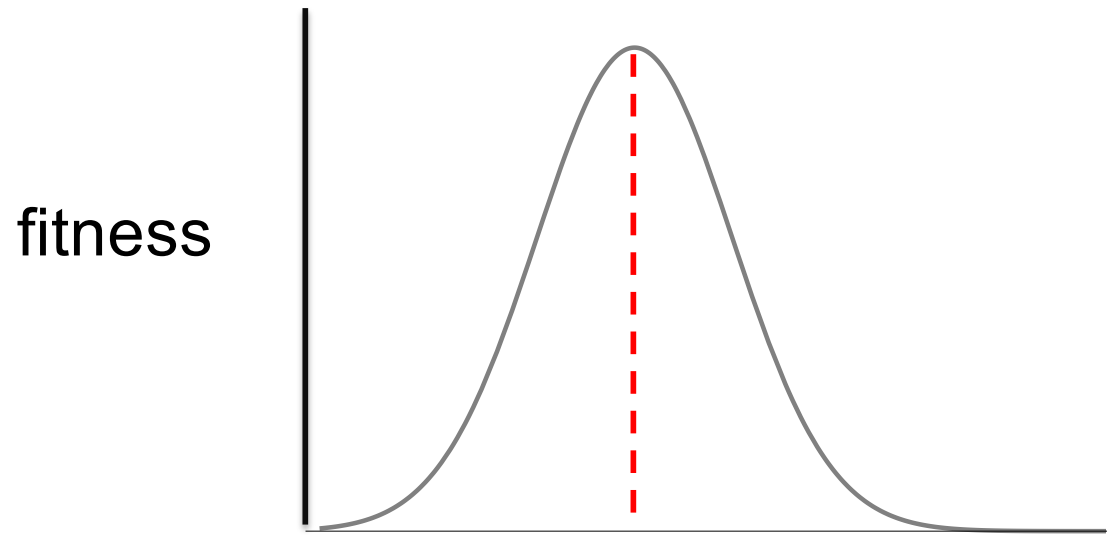
tracking environmental change



tracking environmental change



tracking environmental change

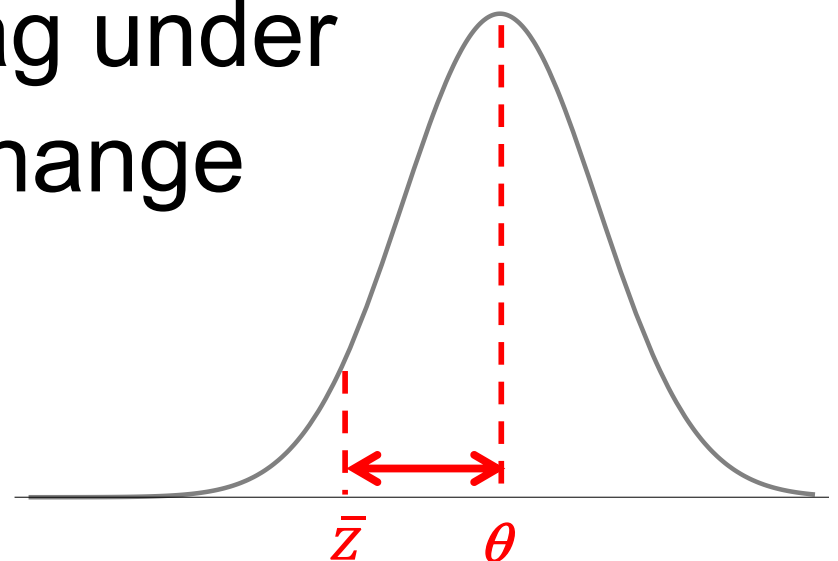


evolutionary lag

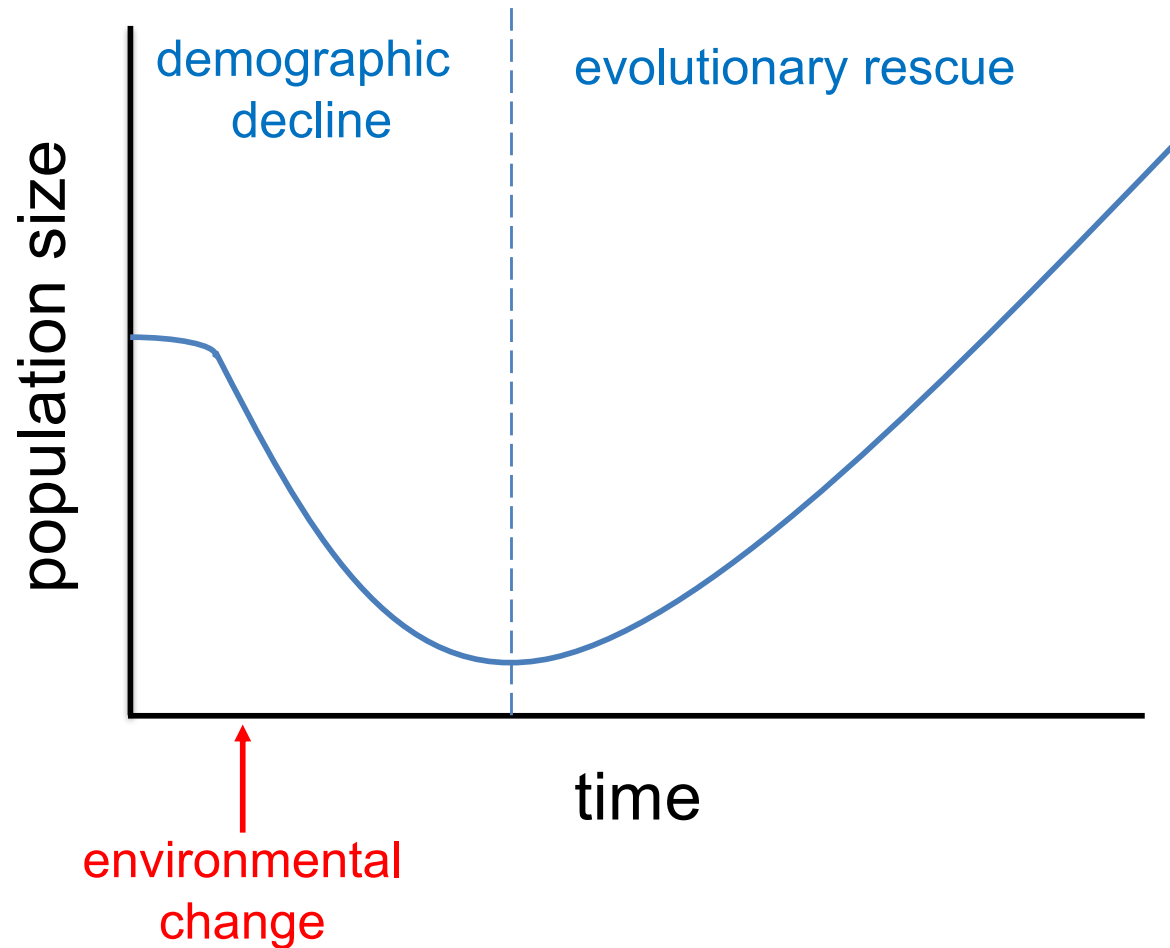
- evolutionary lag (or lag load)
 - difference between phenotypic trait mean and its optimum

$$L_{\theta} = \frac{(\bar{z} - \theta)^2}{2\sigma_w^2} \quad \text{Maynard Smith (1976)}$$

- greater evolutionary lag under rapid environmental change

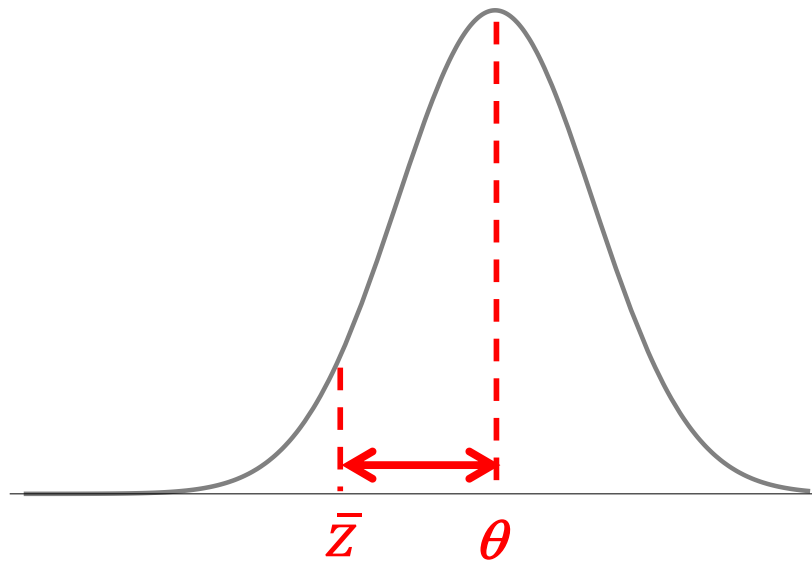


evolutionary rescue



complex life histories and adaptation

- how do stage structure and clonal reproduction affect a population's ability to track environmental change?



stage structure and clonality



image by Forest & Kim Starr.



image by Nadiatalent



image from Oxford Scientific



image from NOAA website

Life-history complexities

- mutation arising in somatic tissues
 - in gametes and/or independent clonal offspring
 - within-individual or somatic selection

somatic mutation

gametic
reproduction



Image: Jouko Lehmuskallio

agametic (clonal)
reproduction



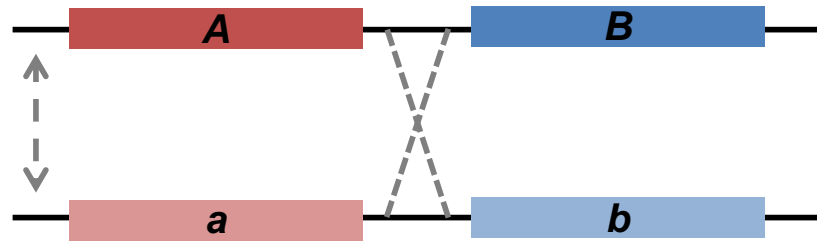
image from Oxford Scientific

Life-history complexities

- mutation arising in somatic tissues
 - in gametes and/or independent clonal offspring
 - within-individual or somatic selection
- reproduction without meiosis
 - shields from higher meiotic mutation rates
 - lacks genetic segregation (heterozygosity, homozygosity)
 - lacks recombination

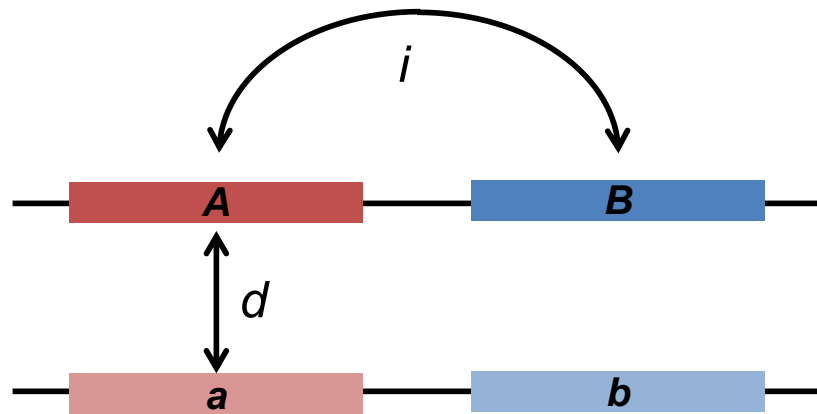
sexual reproduction

α



clonal reproduction

α, d, i



Life-history complexities

- mutation arising in somatic tissues
 - in gametes and/or independent clonal offspring
 - within-individual or somatic selection
- reproduction without meiosis
 - shields from higher meiotic mutation rates
 - lacks genetic segregation (heterozygosity, homozygosity)
 - lacks recombination
- clonal offspring phenotypically distinct from sexual offspring

clonal reproduction and invasive spread

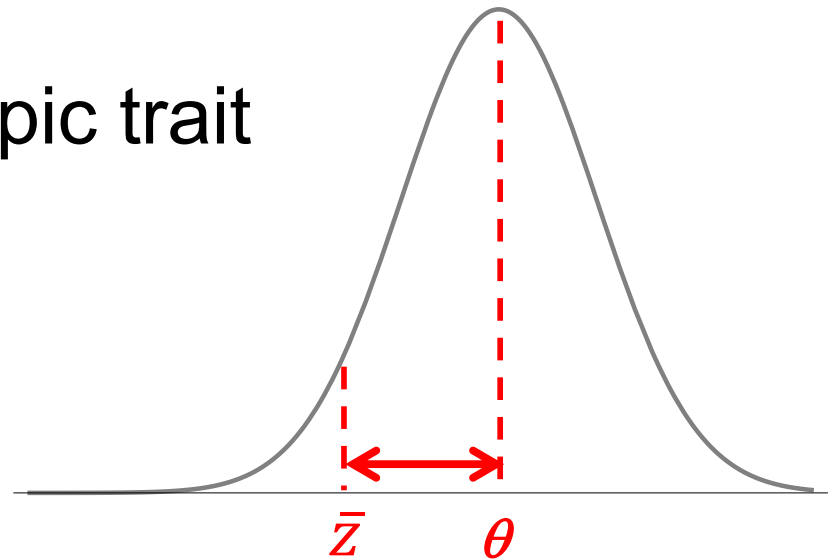


phenotypic evolution with stage-structured life histories

- multivariate phenotypic trait

$$\mathbf{z} = (z_1, z_2, \dots, z_n)^T$$

$$\mathbf{z} = \mathbf{g} + \mathbf{e}$$



- N_i = number of individuals for each stage i
- $\bar{\mathbf{g}}_i$ = mean genotype of stage i
- $\bar{\mathbf{z}}_i$ = mean phenotype of stage i

dynamical models: describing systems that change over time

differential equations – describe the *rate* at which
a variable changes over time;
continuous in time

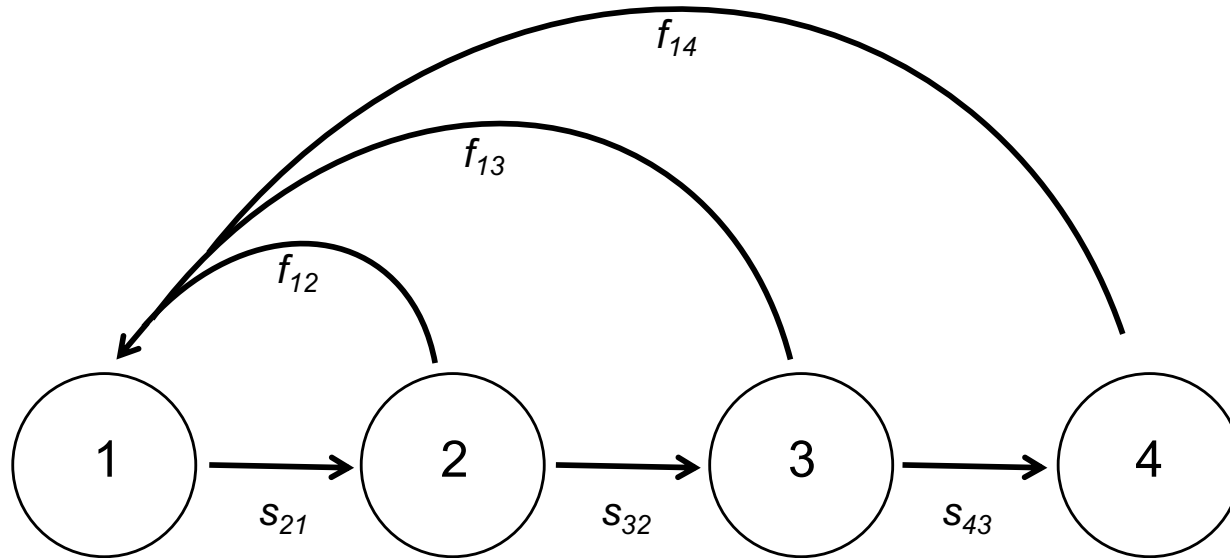
$$\frac{d n(t)}{dt} = \text{"some function of } n(t)\text{"}$$

recursion equations – describe the *value* of a variable
in the next time step;
discrete in time

$$n(t + 1) = \text{"some function of } n(t)\text{"}$$

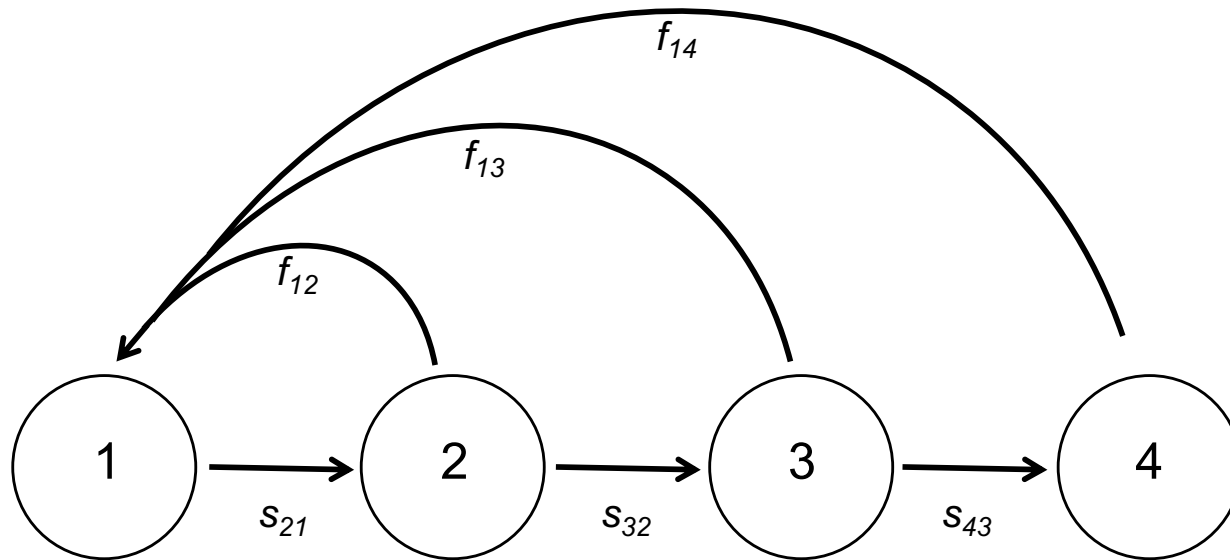
$$n' = \text{"some function of } n\text{"}$$

age-structured life history graph



$$N' = \mathbf{A} N \quad \begin{bmatrix} N_1 \\ N_2 \\ N_3 \\ N_4 \end{bmatrix}' = \begin{bmatrix} 0 & f_{12} & f_{13} & f_{14} \\ s_{21} & 0 & 0 & 0 \\ 0 & s_{32} & 0 & 0 \\ 0 & 0 & s_{43} & 0 \end{bmatrix} \begin{bmatrix} N_1 \\ N_2 \\ N_3 \\ N_4 \end{bmatrix}$$

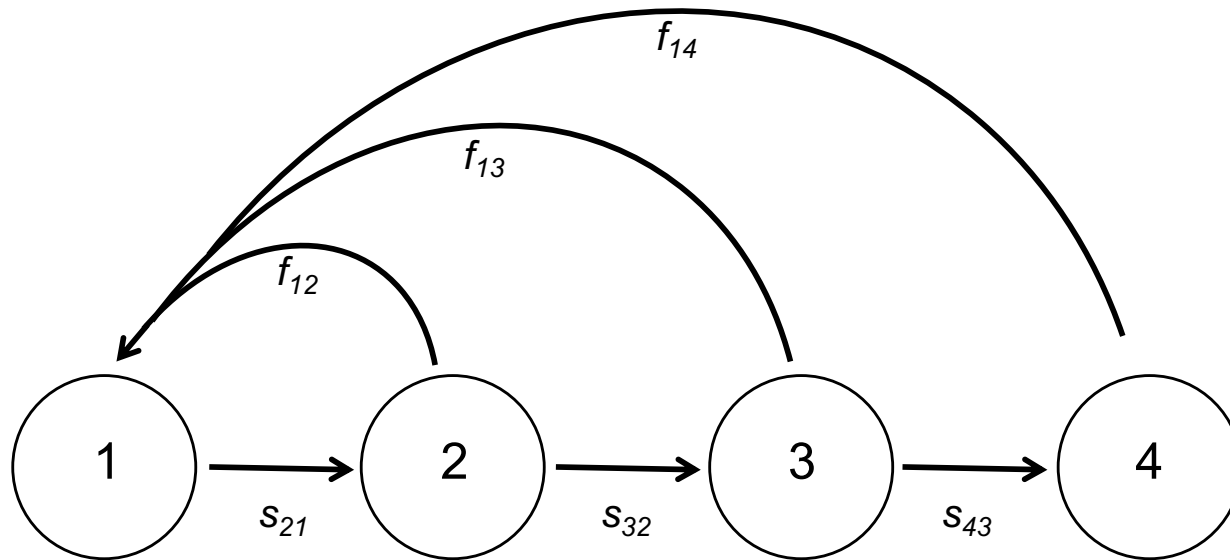
age-structured life history graph



$$N' = \mathbf{A} N \quad \begin{bmatrix} N_1' \\ N_2 \\ N_3 \\ N_4 \end{bmatrix} = \begin{bmatrix} 0 & f_{12} & f_{13} & f_{14} \\ s_{21} & 0 & 0 & 0 \\ 0 & s_{32} & 0 & 0 \\ 0 & 0 & s_{43} & 0 \end{bmatrix} \begin{bmatrix} N_1 \\ N_2 \\ N_3 \\ N_4 \end{bmatrix}$$

$$N_1' = f_{12}N_2 + f_{13}N_3 + f_{14}N_4$$

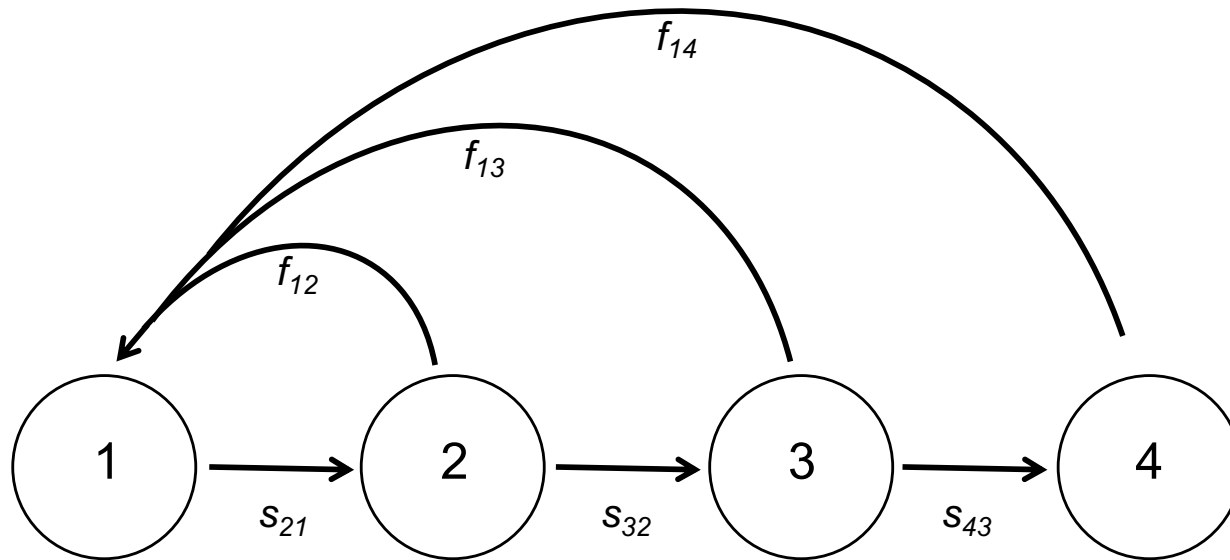
age-structured life history graph



$$N' = \mathbf{A} N \quad \begin{bmatrix} N_1' \\ N_2' \\ N_3' \\ N_4' \end{bmatrix} = \begin{bmatrix} 0 & f_{12} & f_{13} & f_{14} \\ s_{21} & 0 & 0 & 0 \\ 0 & s_{32} & 0 & 0 \\ 0 & 0 & s_{43} & 0 \end{bmatrix} \begin{bmatrix} N_1 \\ N_2 \\ N_3 \\ N_4 \end{bmatrix}$$

$$N_2' = s_{21} N_1$$

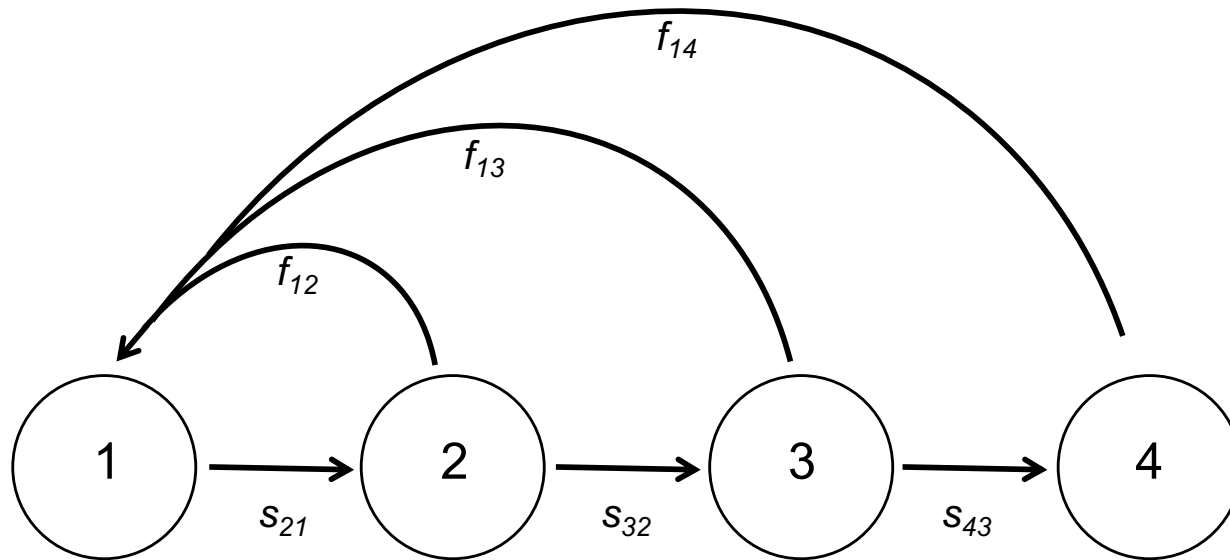
age-structured life history graph



$$N' = A N \quad \begin{bmatrix} N_1' \\ N_2 \\ N_3 \\ N_4 \end{bmatrix} = \begin{bmatrix} 0 & f_{12} & f_{13} & f_{14} \\ s_{21} & 0 & 0 & 0 \\ 0 & s_{32} & 0 & 0 \\ 0 & 0 & s_{43} & 0 \end{bmatrix} \begin{bmatrix} N_1 \\ N_2 \\ N_3 \\ N_4 \end{bmatrix}$$

$$N_3' = s_{32} N_2$$

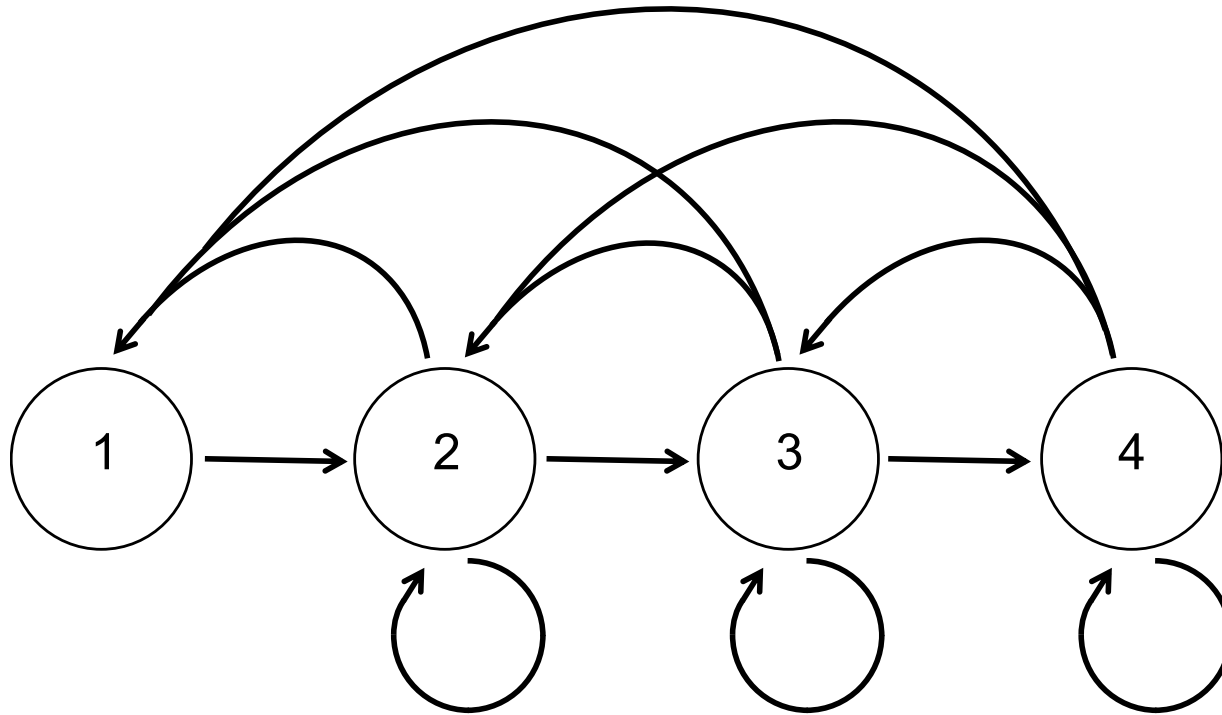
age-structured life history graph



$$N' = \mathbf{A} N \quad \begin{bmatrix} N_1' \\ N_2' \\ N_3' \\ N_4' \end{bmatrix} = \begin{bmatrix} 0 & f_{12} & f_{13} & f_{14} \\ s_{21} & 0 & 0 & 0 \\ 0 & s_{32} & 0 & 0 \\ 0 & 0 & s_{43} & 0 \end{bmatrix} \begin{bmatrix} N_1 \\ N_2 \\ N_3 \\ N_4 \end{bmatrix}$$

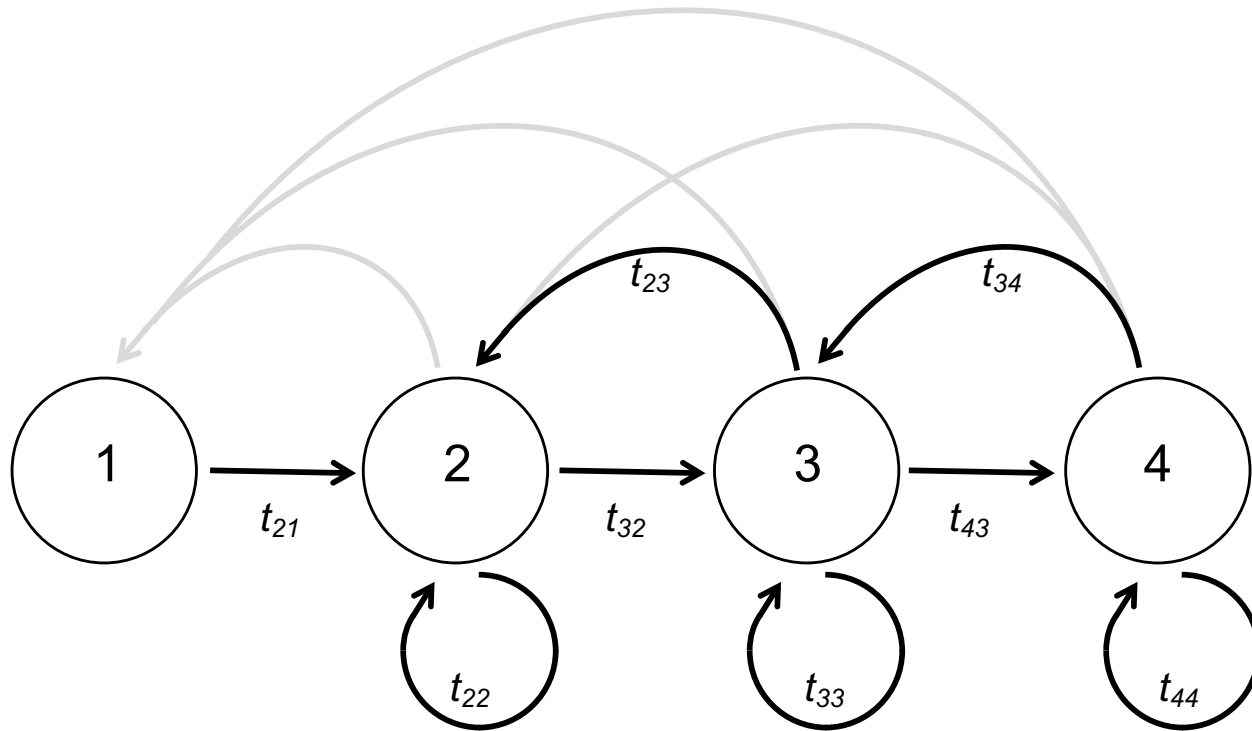
$$N_4' = s_{43} N_3$$

stage-structured life history graph



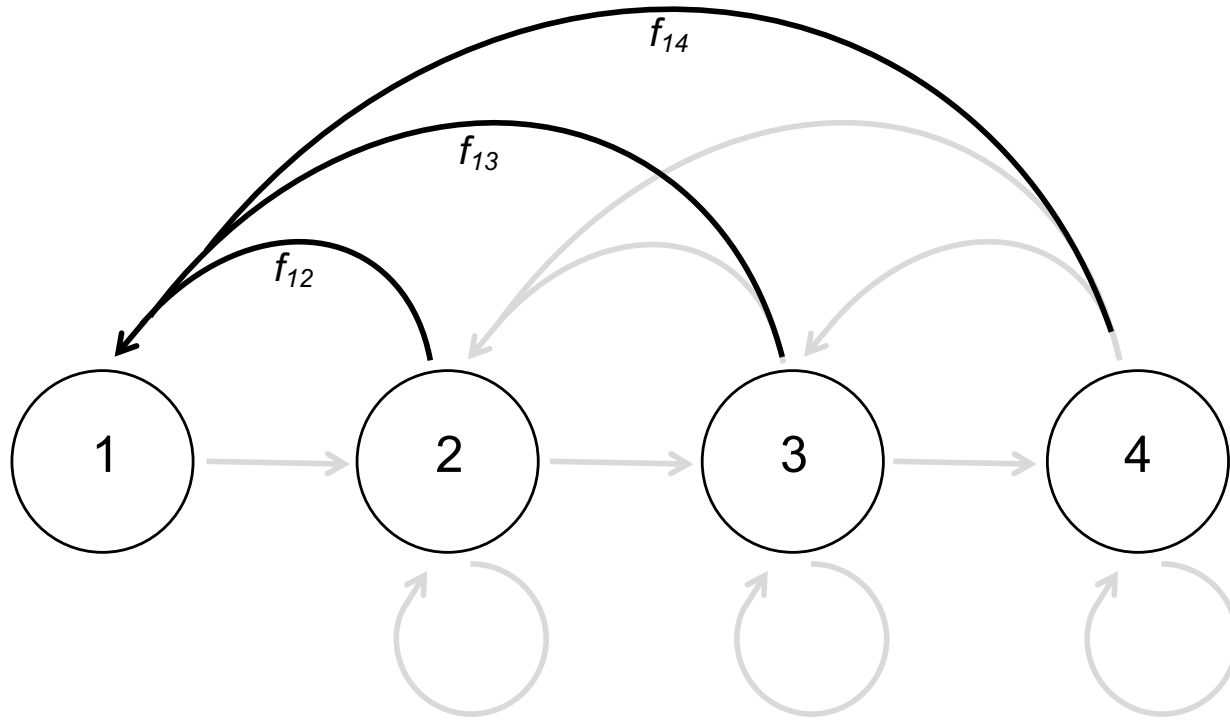
$$N' = \mathbf{A} N \quad \mathbf{A} = \begin{bmatrix} 0 & f_{12} & f_{13} & f_{14} \\ t_{21} & t_{22} & t_{23} + c_{23} & c_{24} \\ 0 & t_{32} & t_{33} & t_{34} \\ 0 & 0 & t_{43} & t_{44} \end{bmatrix} \quad N = \begin{bmatrix} N_1 \\ N_2 \\ N_3 \\ N_4 \end{bmatrix}$$

transitions



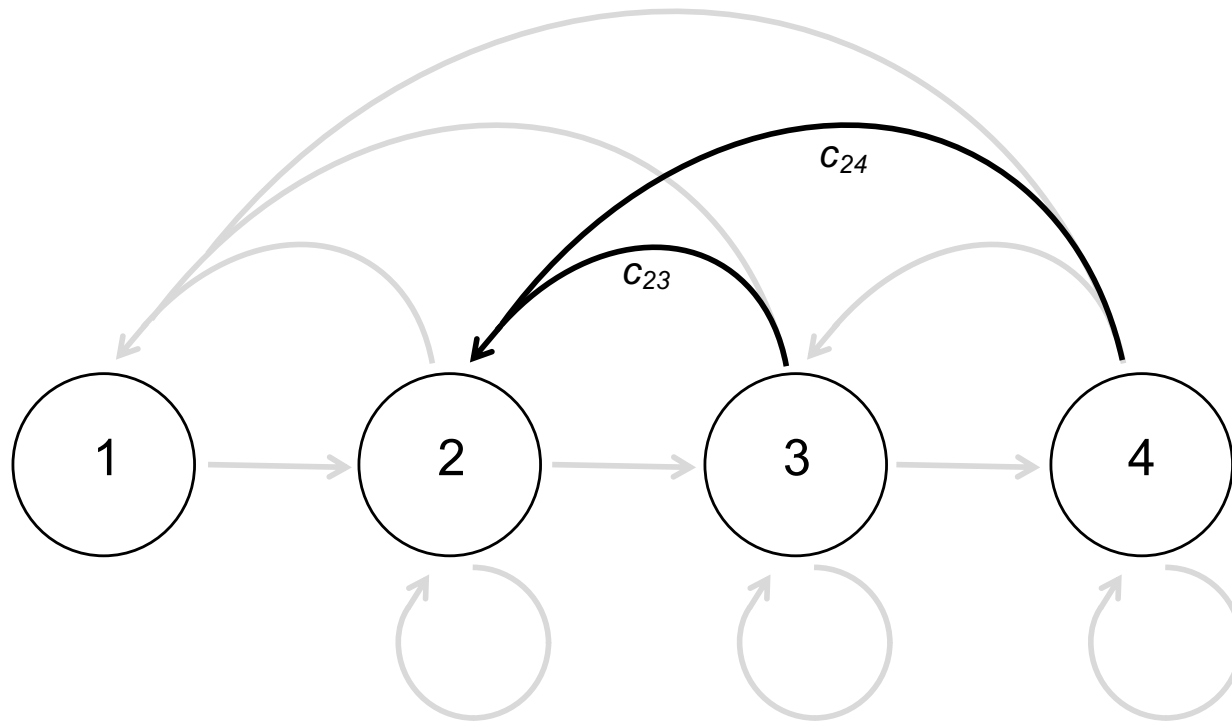
$$\begin{bmatrix} 0 & f_{12} & f_{13} & f_{14} \\ t_{21} & t_{22} & t_{23} + c_{23} & c_{24} \\ 0 & t_{32} & t_{33} & t_{34} \\ 0 & 0 & t_{43} & t_{44} \end{bmatrix}$$

sexual reproduction



$$\begin{bmatrix} 0 & f_{12} & f_{13} & f_{14} \\ t_{21} & t_{22} & t_{23} + c_{23} & c_{24} \\ 0 & t_{32} & t_{33} & t_{34} \\ 0 & 0 & t_{43} & t_{44} \end{bmatrix}$$

clonal reproduction



$$\begin{bmatrix} 0 & f_{12} & f_{13} & f_{14} \\ t_{21} & t_{22} & t_{23} + c_{23} & c_{24} \\ 0 & t_{32} & t_{33} & t_{34} \\ 0 & 0 & t_{43} & t_{44} \end{bmatrix}$$

Explicitly considering clonal reproduction

- three types of movements

$$\begin{aligned} N'_i &= \sum_j N_j \bar{a}_{ij} = \sum_j N_j (\bar{t}_{ij} + \bar{f}_{ij} + \bar{c}_{ij}) \\ &= \sum_j N_j \bar{t}_{ij} + \sum_j N_j \bar{f}_{ij} + \sum_j N_j \bar{c}_{ij} = T'_i + F'_i + C'_i \end{aligned}$$

t_{ij} = transition from stage j to stage i

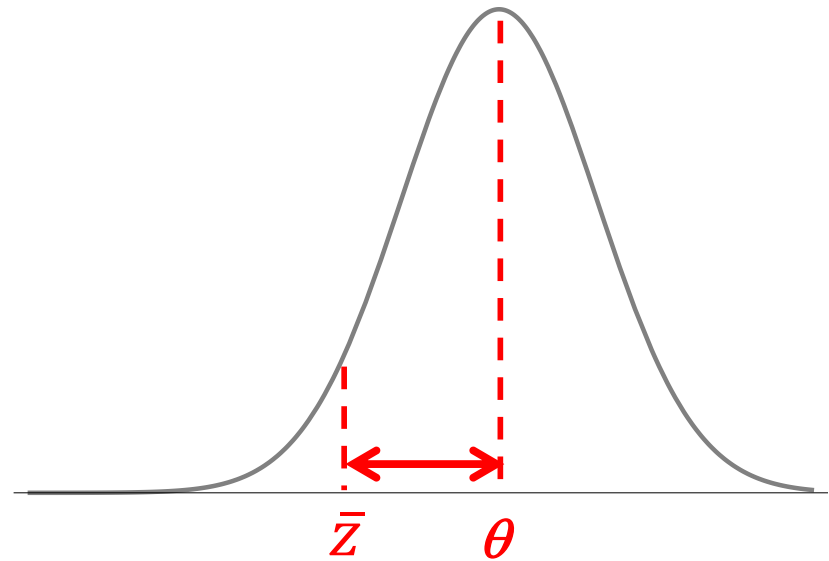
f_{ij} = sexual reproduction from stage j to stage i

c_{ij} = clonal reproduction from stage j to stage i

phenotypic evolution

z = phenotypic trait

$$\mathbf{z} = \mathbf{g} + \mathbf{e}$$



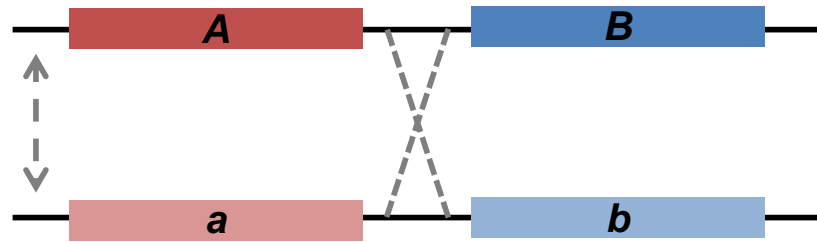
g = additive genetic factor

e = non-additive genetic
+ random environmental factor

What makes up e ?

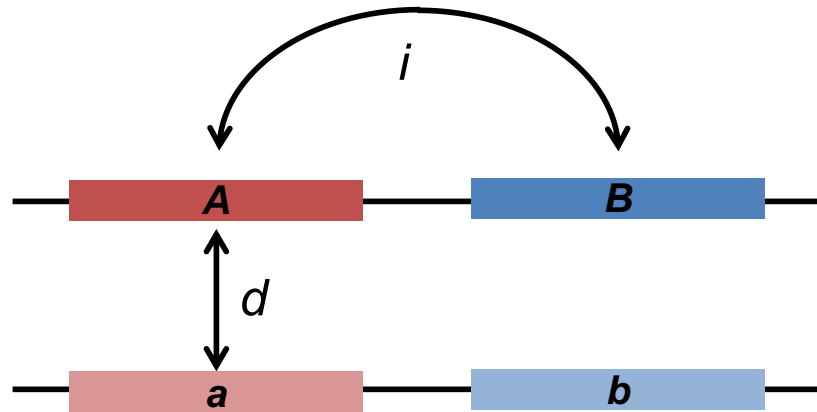
sexual reproduction

α



clonal reproduction

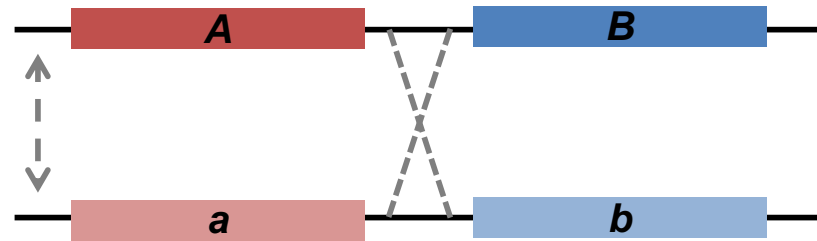
α, d, i



What makes up e ?

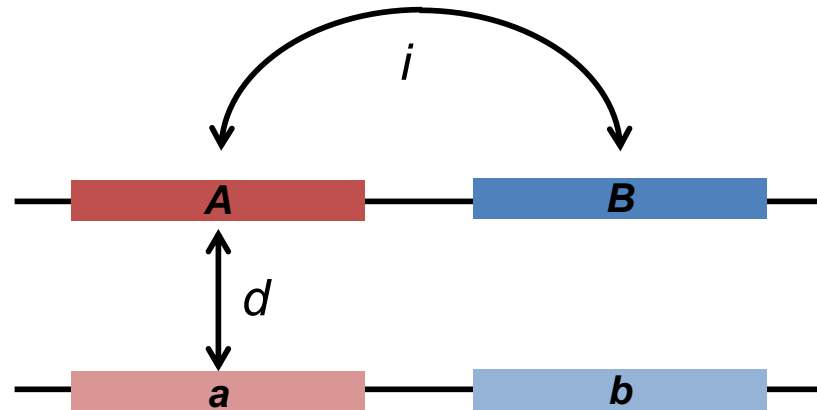
sexual reproduction

α



clonal reproduction

α, d, i

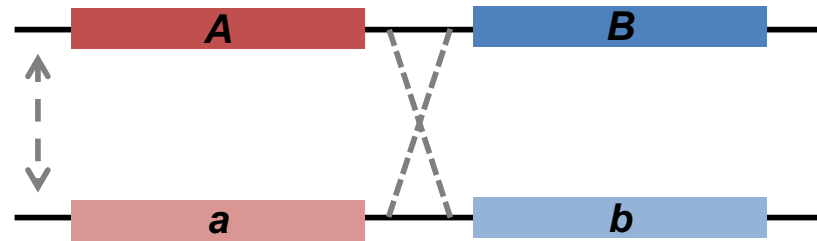


non-additive genetic factors
(dominance, epistasis)

What makes up e ?

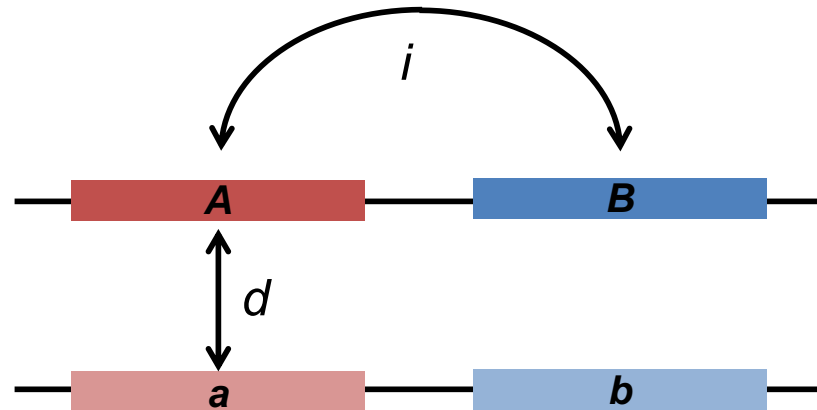
sexual reproduction

α



clonal reproduction

α, d, i



non-additive genetic factors
(dominance, epistasis)

+ random environmental factors

ρ = association between \mathbf{e} in parent and clonal offspring

ρ close to 0



Joerg Hauke/Getty Images

ρ close to 1



© Steffen Claus

recursions for phenotypic and genotypic means

$$\bar{\mathbf{z}}'_i = \sum_j \left[(d_{ij}^t + \mathbf{R}d_{ij}^c) \bar{\mathbf{z}}_j + (d_{ij}^f + (\mathbf{I} - \mathbf{R})d_{ij}^c) \bar{\mathbf{g}}_j + \frac{d_{ij}}{\bar{a}_{ij}} \left(\mathbf{P}_j \nabla_{\bar{\mathbf{z}}_j} \bar{t}_{ij} + \mathbf{G}_j \nabla_{\bar{\mathbf{z}}_j} \bar{f}_{ij} + \mathbf{R} \mathbf{P}_j \nabla_{\bar{\mathbf{z}}_j} \bar{c}_{ij} + (\mathbf{I} - \mathbf{R}) \mathbf{G}_j \nabla_{\bar{\mathbf{z}}_j} \bar{c}_{ij} \right) \right]$$

$$\bar{\mathbf{g}}'_i = \sum_j d_{ij} \bar{\mathbf{g}}_j + \sum_j d_{ij} \mathbf{G}_j \nabla_{\bar{\mathbf{z}}_j} \ln \bar{a}_{ij}$$

\mathbf{P}_j = phenotypic covariance matrix

$$\nabla_{\bar{\mathbf{z}}_j} = (\partial/\partial \bar{z}_1, \partial/\partial \bar{z}_2, \dots, \partial/\partial \bar{z}_m)^T$$

\mathbf{G}_j = additive genetic covariance matrix

$$\mathbf{R} = \begin{bmatrix} \rho_1 & 0 & \dots & 0 \\ 0 & \rho_2 & & 0 \\ \vdots & & \ddots & \vdots \\ 0 & \dots & \dots & \rho_m \end{bmatrix}$$

$$\bar{a}_{ij} = \bar{t}_{ij} + \bar{f}_{ij} + \bar{c}_{ij}$$

$$d_{ij} = \bar{a}_{ij} N_j / N'_i$$

$$d_{ij}^t = \bar{t}_{ij} N_j / N'_i, \quad d_{ij}^f = \bar{f}_{ij} N_j / N'_i, \quad d_{ij}^c = \bar{c}_{ij} N_j / N'_i$$

recursions for phenotypic and genotypic means

$$\bar{\mathbf{z}}'_i = \sum_j \left[(d_{ij}^t + \mathbf{R}d_{ij}^c)\bar{\mathbf{z}}_j + (d_{ij}^f + (\mathbf{I} - \mathbf{R})d_{ij}^c)\bar{\mathbf{g}}_j + \frac{d_{ij}}{\bar{a}_{ij}} \left(\mathbf{P}_j \nabla_{\bar{\mathbf{z}}_j} \bar{t}_{ij} + \mathbf{G}_j \nabla_{\bar{\mathbf{z}}_j} \bar{f}_{ij} + \mathbf{R} \mathbf{P}_j \nabla_{\bar{\mathbf{z}}_j} \bar{c}_{ij} + (\mathbf{I} - \mathbf{R}) \mathbf{G}_j \nabla_{\bar{\mathbf{z}}_j} \bar{c}_{ij} \right) \right]$$

$$\bar{\mathbf{g}}'_i = \sum_j d_{ij} \bar{\mathbf{g}}_j + \sum_j d_{ij} \mathbf{G}_j \nabla_{\bar{\mathbf{z}}_j} \ln \bar{a}_{ij}$$

\mathbf{P}_j = phenotypic covariance matrix

$$\nabla_{\bar{\mathbf{z}}_j} = (\partial/\partial \bar{z}_1, \partial/\partial \bar{z}_2, \dots, \partial/\partial \bar{z}_m)^T$$

\mathbf{G}_j = additive genetic covariance matrix

$$\mathbf{R} = \begin{bmatrix} \rho_1 & 0 & \dots & 0 \\ 0 & \rho_2 & & 0 \\ \vdots & & \ddots & \vdots \\ 0 & \dots & \dots & \rho_m \end{bmatrix}$$

$$\bar{a}_{ij} = \bar{t}_{ij} + \bar{f}_{ij} + \bar{c}_{ij}$$

$$d_{ij} = \bar{a}_{ij} N_j / N'_i$$

$$d_{ij}^t = \bar{t}_{ij} N_j / N'_i, \quad d_{ij}^f = \bar{f}_{ij} N_j / N'_i, \quad d_{ij}^c = \bar{c}_{ij} N_j / N'_i$$

recursions for phenotypic and genotypic means

$$\bar{\mathbf{z}}'_i = \sum_j \left[(d_{ij}^t + \mathbf{R}d_{ij}^c) \bar{\mathbf{z}}_j + (d_{ij}^f + (\mathbf{I} - \mathbf{R})d_{ij}^c) \bar{\mathbf{g}}_j + \frac{d_{ij}}{\bar{a}_{ij}} \left(\mathbf{P}_j \nabla_{\bar{\mathbf{z}}_j} \bar{t}_{ij} + \mathbf{G}_j \nabla_{\bar{\mathbf{z}}_j} \bar{f}_{ij} + \mathbf{R} \mathbf{P}_j \nabla_{\bar{\mathbf{z}}_j} \bar{c}_{ij} + (\mathbf{I} - \mathbf{R}) \mathbf{G}_j \nabla_{\bar{\mathbf{z}}_j} \bar{c}_{ij} \right) \right]$$

$$\bar{\mathbf{g}}'_i = \sum_j d_{ij} \bar{\mathbf{g}}_j + \sum_j d_{ij} \mathbf{G}_j \nabla_{\bar{\mathbf{z}}_j} \ln \bar{a}_{ij}$$

\mathbf{P}_j = phenotypic covariance matrix

$$\nabla_{\bar{\mathbf{z}}_j} = (\partial/\partial \bar{z}_1, \partial/\partial \bar{z}_2, \dots, \partial/\partial \bar{z}_m)^T$$

\mathbf{G}_j = additive genetic covariance matrix

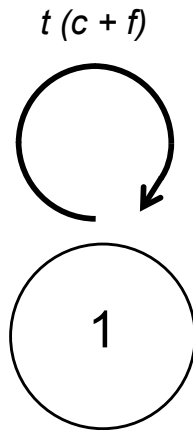
$$\mathbf{R} = \begin{bmatrix} \rho_1 & 0 & \dots & 0 \\ 0 & \rho_2 & & 0 \\ \vdots & & \ddots & \vdots \\ 0 & \dots & \dots & \rho_m \end{bmatrix}$$

$$\bar{a}_{ij} = \bar{t}_{ij} + \bar{f}_{ij} + \bar{c}_{ij}$$

$$d_{ij} = \bar{a}_{ij} N_j / N'_i$$

$$d_{ij}^t = \bar{t}_{ij} N_j / N'_i, \quad d_{ij}^f = \bar{f}_{ij} N_j / N'_i, \quad d_{ij}^c = \bar{c}_{ij} N_j / N'_i$$

Simple life history



clonal and sexual reproduction

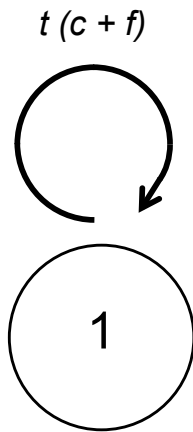
selection on survival probability $t = \exp \left[-\frac{(\bar{z}_1 - \theta)^2}{(2\omega^2)} \right]$

amount of clonal reproduction $r_c = c/(f + c)$

$$\Delta \bar{z} = (1 - r_c \rho)(\bar{g} - \bar{z}) + \{r_c \rho P + (1 - r_c \rho)G\} \frac{\theta - \bar{z}}{\omega^2}$$

$$\Delta \bar{g} = G \frac{\theta - \bar{z}}{\omega^2}$$

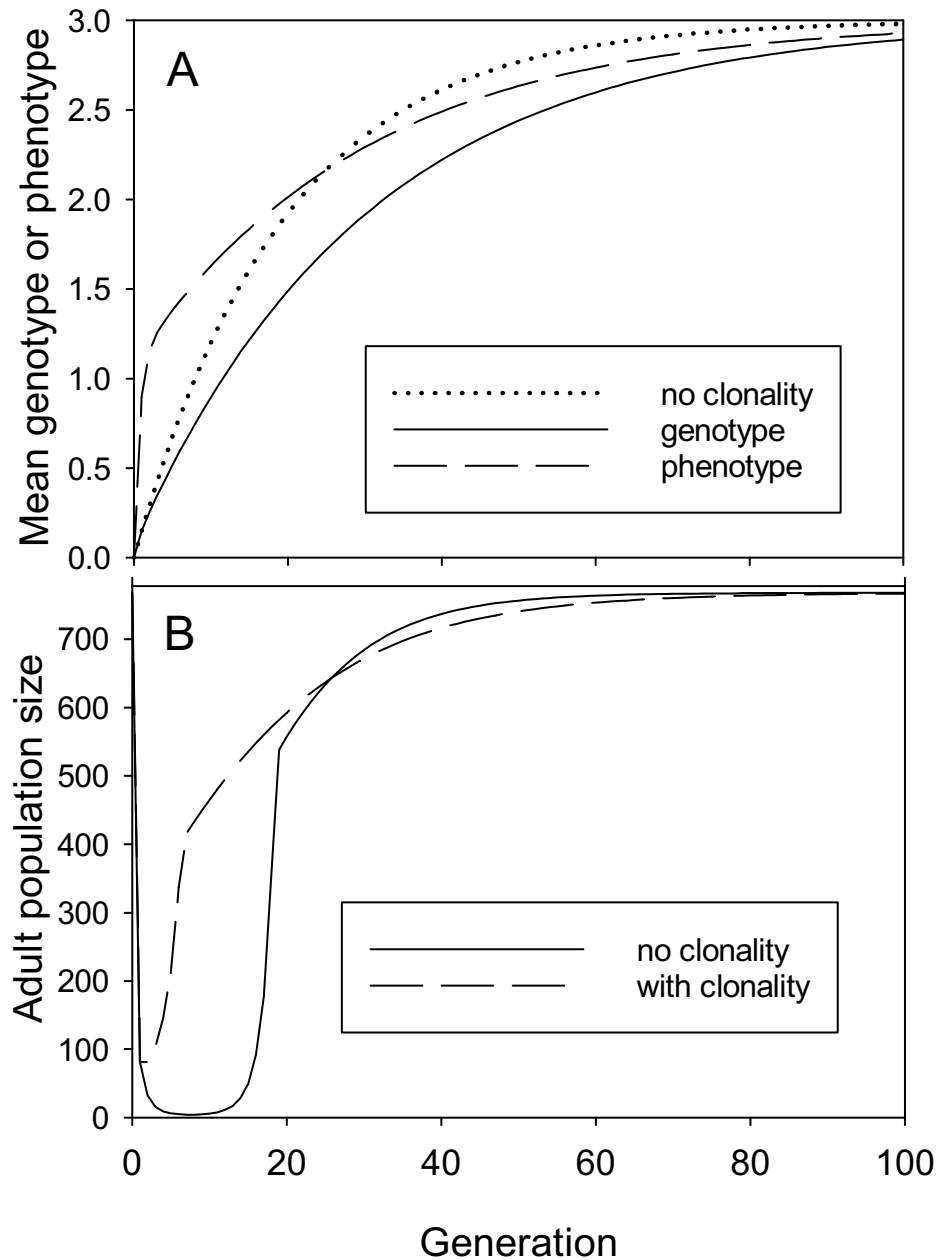
analytical results – effect of clonality



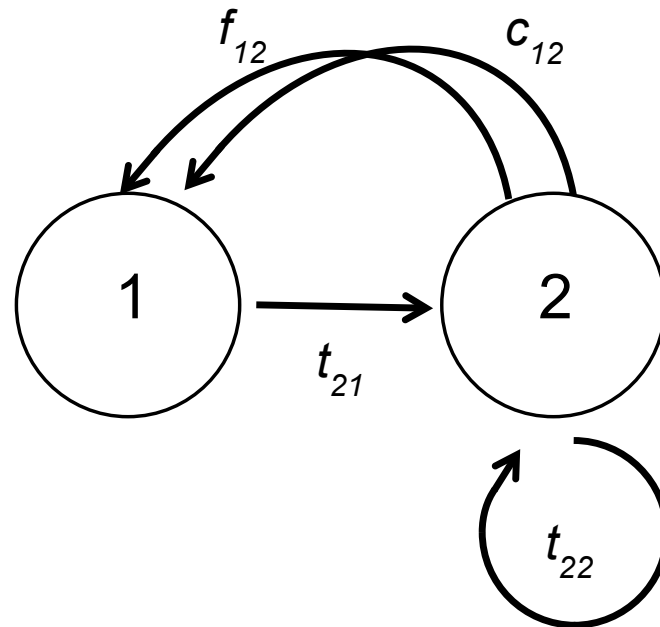
no clonality ($r_c \rho = 0$)

vs.

with clonality ($r_c \rho = 0.5$)

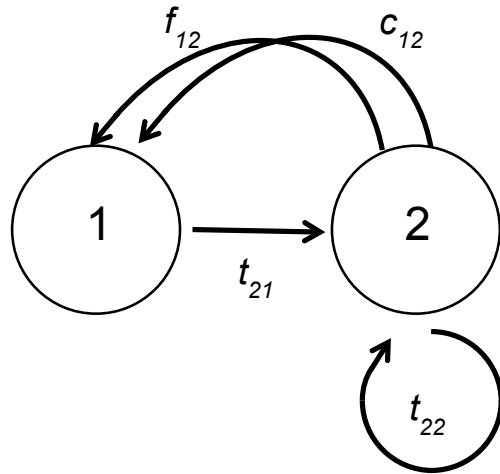


Life history with stage structure



$$\begin{pmatrix} N_1 \\ N_2 \end{pmatrix}' = \begin{pmatrix} 0 & c_{12} + f_{12} \\ t_{21} & t_{22} \end{pmatrix} \begin{pmatrix} N_1 \\ N_2 \end{pmatrix}$$

Life history with stage structure



selection on juvenile survival

$$\bar{t}_{21} = t_{\max} \exp \left[-\frac{(\bar{z}_1 - \theta)^2}{(2\omega^2)} \right]$$

$$r_c = c_{12} / (c_{12} + f_{12})$$

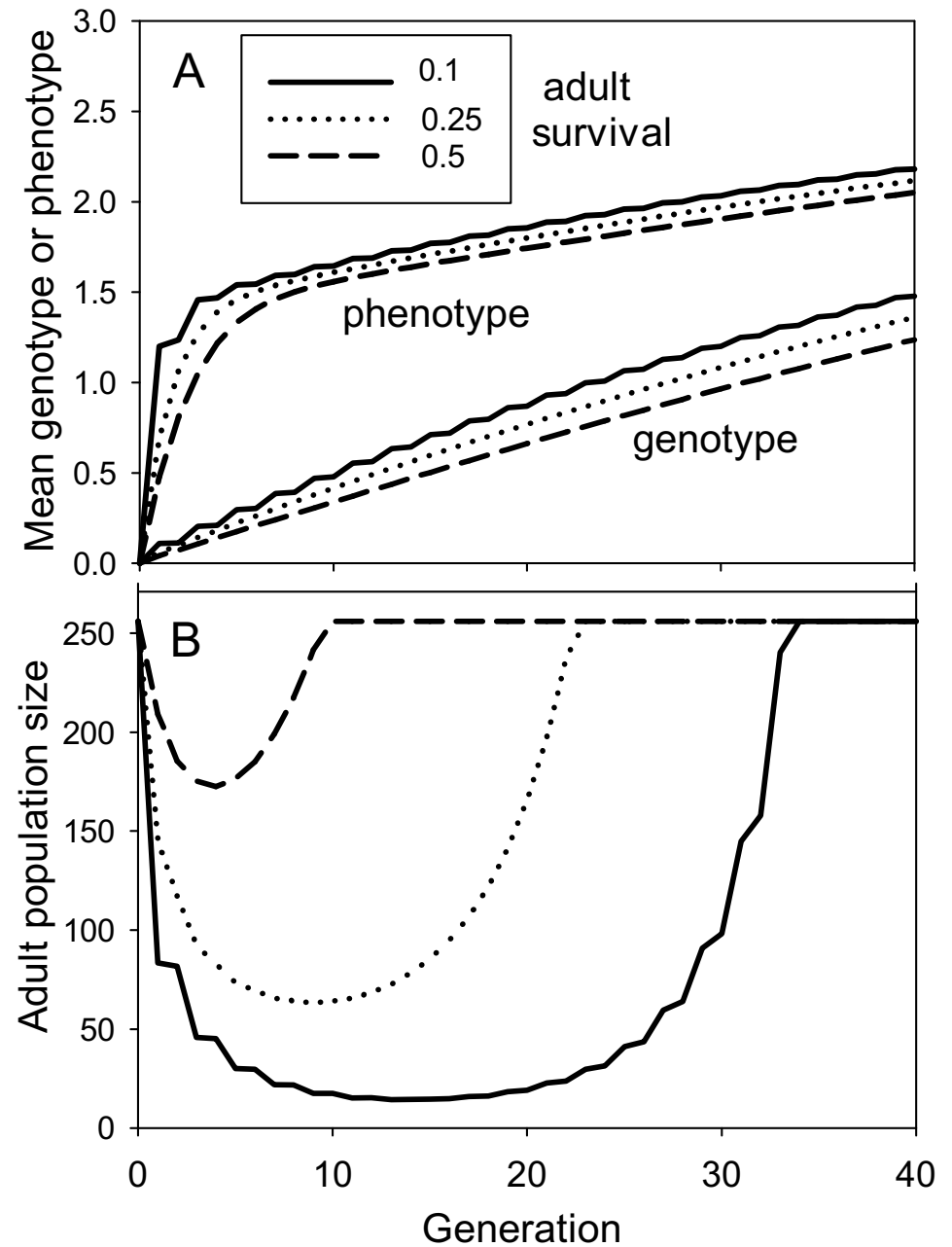
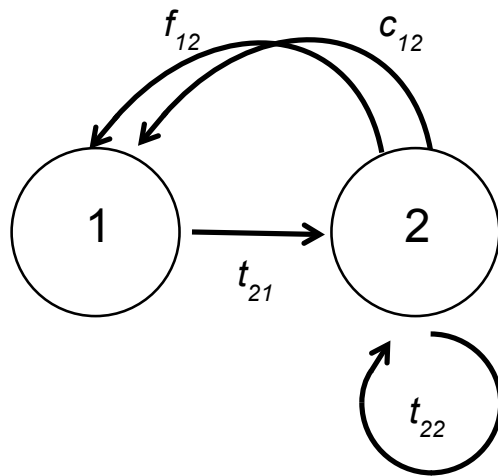
$$\bar{z}'_1 = r_c \rho (\bar{z}_2 - \bar{g}_2) + \bar{g}_2$$

$$\bar{z}'_2 = \bar{z}_2 + \frac{N_1}{\bar{t}_{21}N_1 + \bar{t}_{22}N_2} \bar{t}_{21} \left[(\bar{z}_1 - \bar{z}_2) + P_1 \left(\frac{\theta - \bar{z}_1}{\omega^2} \right) \right]$$

$$\bar{g}'_1 = \bar{g}_2$$

$$\bar{g}'_2 = \bar{g}_2 + \frac{N_1}{\bar{t}_{21}N_1 + \bar{t}_{22}N_2} \bar{t}_{21} \left[(\bar{g}_1 - \bar{g}_2) + G_1 \left(\frac{\theta - \bar{z}_1}{\omega^2} \right) \right]$$

analytical results – increased adult survival



evolutionary lag in clonal organisms

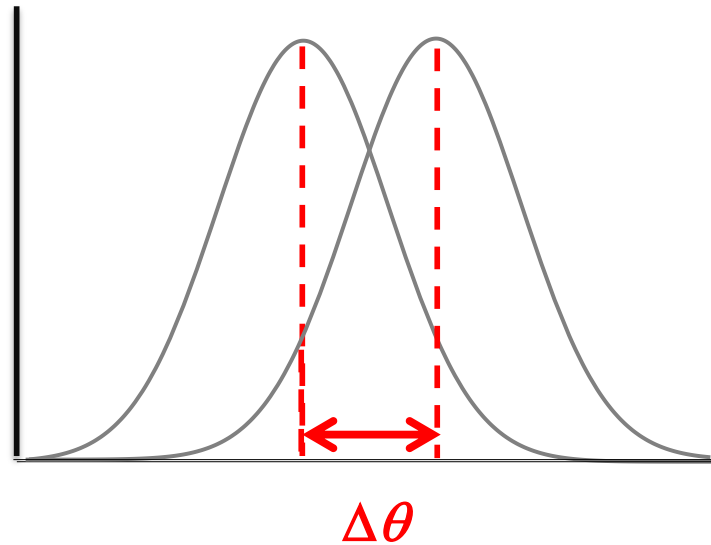
- analytical results
 - clonality ($r_c \rho > 0$) and adult survival (stage structure) both slow approach to equilibrium phenotype
 - but both also reduce both extent and duration of population size decrease
 - demographic advantage

individual-based simulations

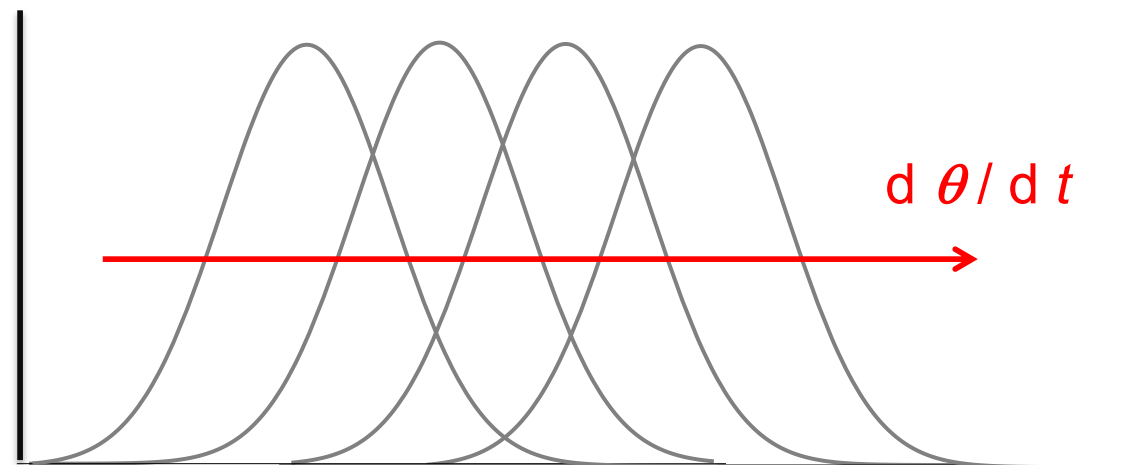
- single polygenic trait z
 - $n = 10$ loci, additive allelic effects
- e normally distributed, mean 0, variance 1
- $\mu_g = 100\mu_s$
- population size ceiling, K
- relative amounts of clonal reproduction, $r_c = c/(c + f)$
- association parameter, ρ
- change in optimum phenotype
 - one-step change
 - continuous, linear change

change in optimum phenotype

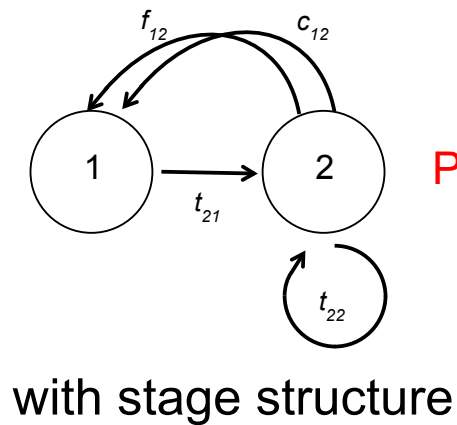
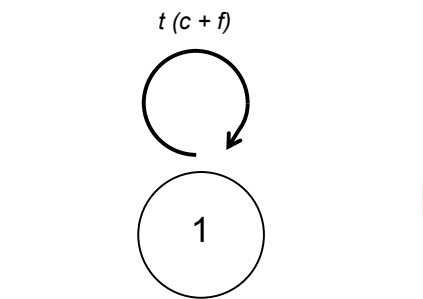
one-step change



continuous change



one-step change in optimal phenotype

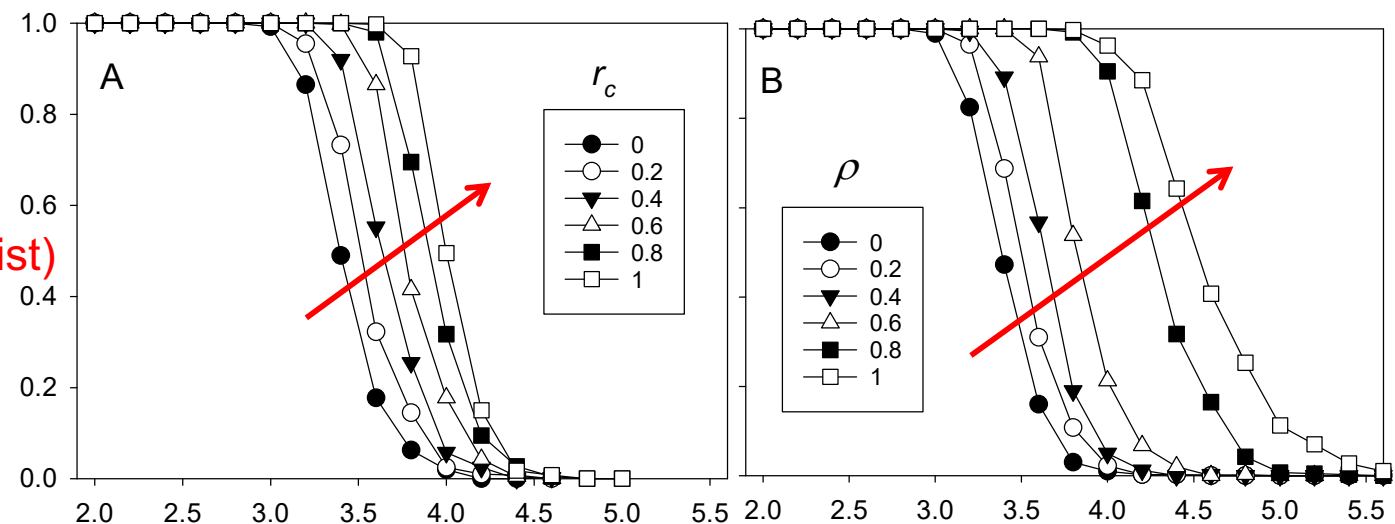
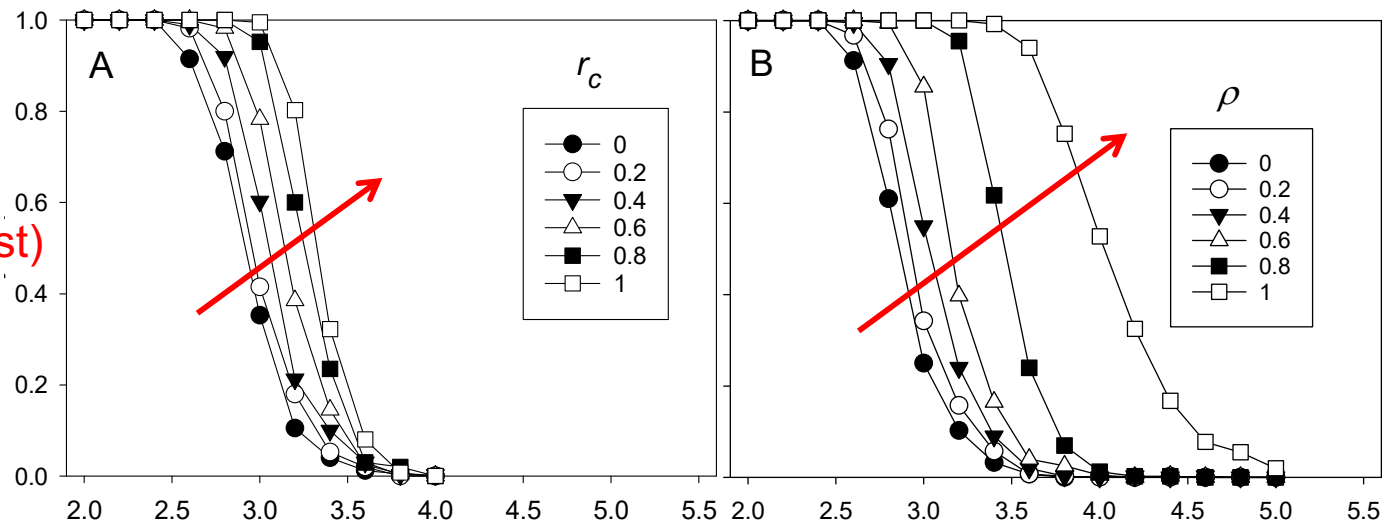


Pr(persist)

Pr(persist)

clonal reproduction r_c
 $\rho = 0.5$

association parameter ρ
 $r_c = 0.5$



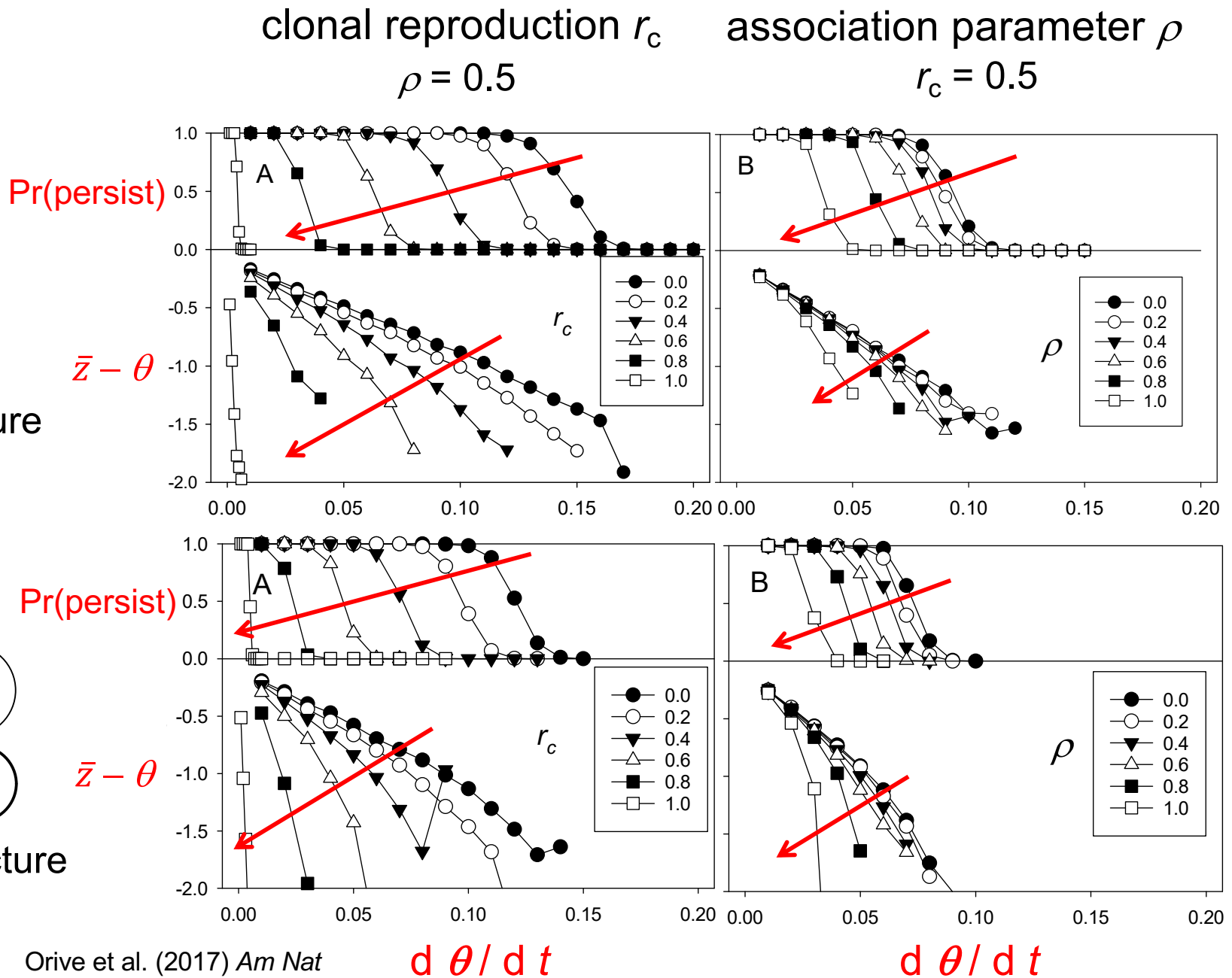
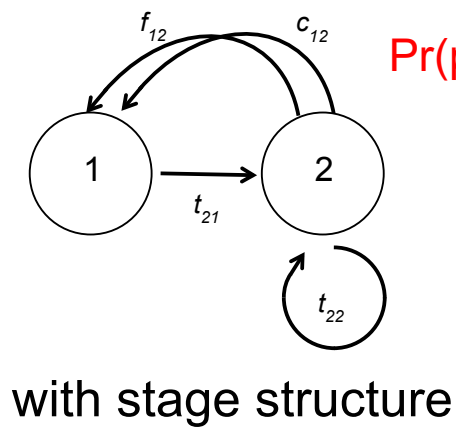
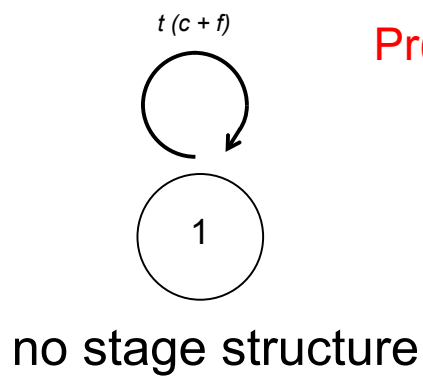
one-step change in optimal phenotype

- greater population persistence with more clonal reproduction (r_c) and higher environmental component association (ρ)
 - standing genotypic variation

one-step change in optimal phenotype

- greater population persistence with more clonal reproduction (r_c) and higher environmental component association (ρ)
 - standing genotypic variation
- stage structure increases probability of population persistence
 - demographic advantage

continuous change in optimal phenotype



continuous, linear change in optimal phenotype

- decreased persistence and greater lag with more clonal reproduction, higher ρ
 - de novo genotypic variation

continuous, linear change in optimal phenotype

- decreased persistence and greater lag with more clonal reproduction, higher ρ
 - de novo genotypic variation
- stage structure decreases persistence and increases lag
 - increased generation time
 - decreased N_e of component of population experiencing phenotypic selection
 - maladaptive “gene flow through time”

evolutionary lag in clonal organisms

- how will clonal organisms respond under rapid environmental change?

evolutionary lag in clonal organisms

- how will clonal organisms respond under rapid environmental change?
- scale of change – whether population experiences that change as a single transition or not

evolutionary lag in clonal organisms

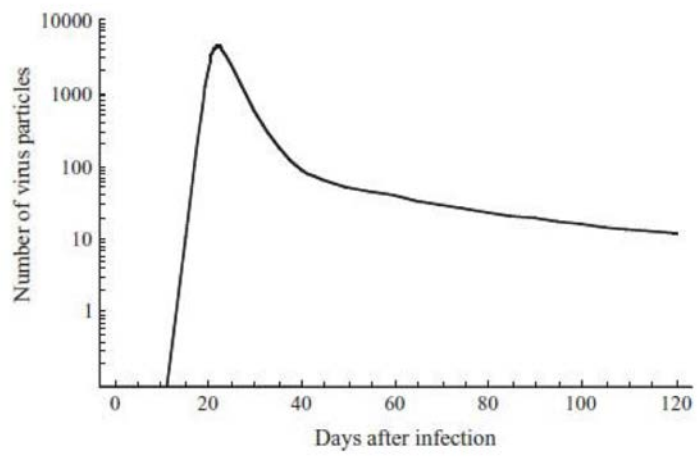
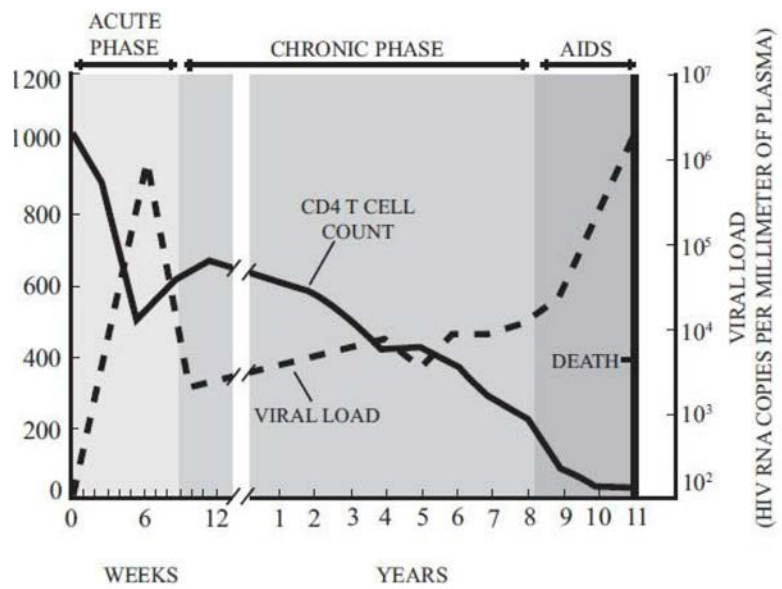
- how will clonal organisms respond under rapid environmental change?
- scale of change – whether population experiences that change as a single transition or not
- amount of phenotypic matching between organisms and their clonal offspring

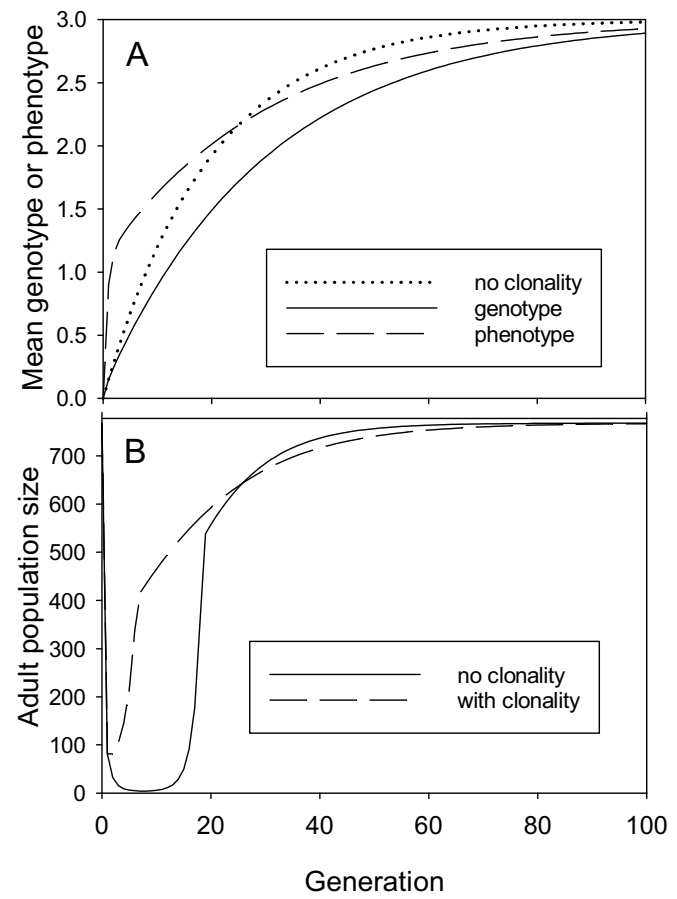
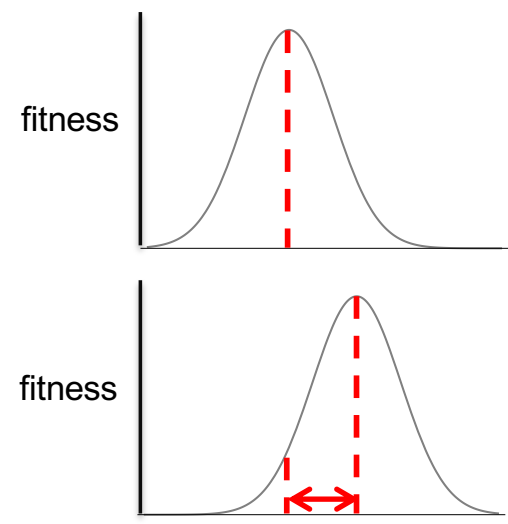
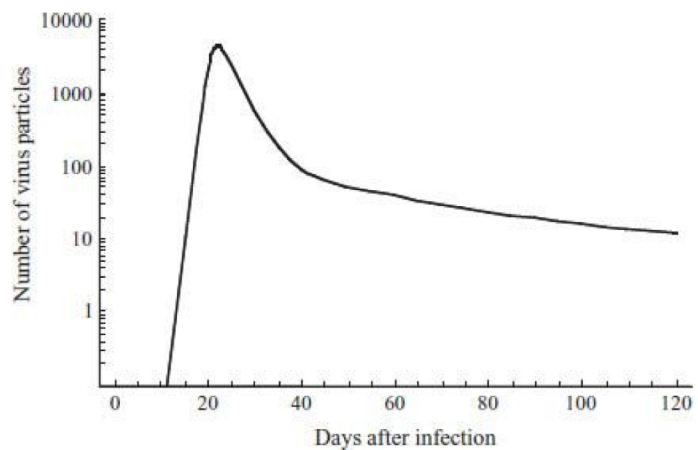
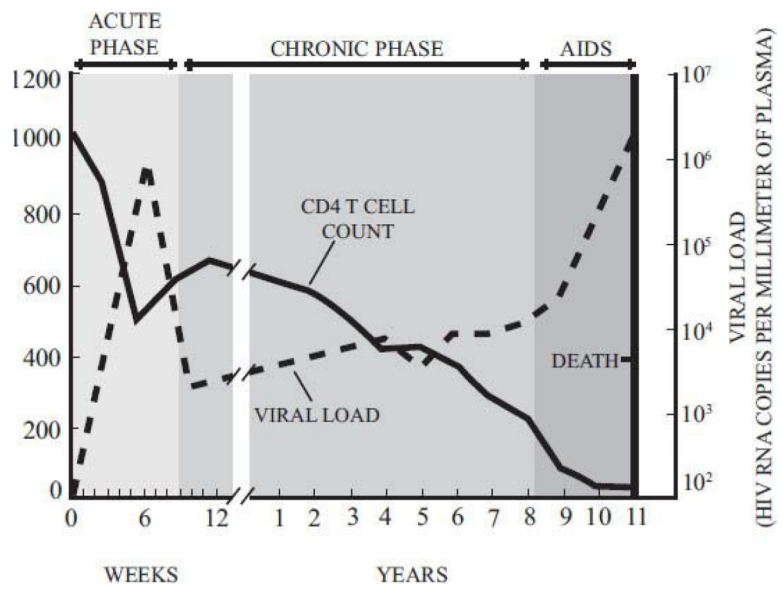
evolutionary lag in clonal organisms

- how will clonal organisms respond under rapid environmental change?
- scale of change – whether population experiences that change as a single transition or not
- amount of phenotypic matching between organisms and their clonal offspring
- existence of stage structured life histories

two ways we can use models to make sense of biology

- Explain what we *do see*
 - Specific test of hypotheses
 - Example: dynamics of HIV after infection
- Predict what we *might see*
 - Generate hypotheses
 - Example: evolutionary lag and rescue with complex life histories





acknowledgments

Orive Lab

Carlos Fernandez

Anna Goddard

Collaborators

Michael Barfield (U Florida)

Bob Holt (U Florida)



KU McNair
Scholars Program

questions?



Website: <http://www.orive.faculty.ku.edu/>

Email: morive@ku.edu

Twitter: [@MEOrive](https://twitter.com/MEOrive)